ORIGINAL ARTICLE

Selenium nanoparticles effect on foot and mouth disease vaccine in local Awassi breed male lambs

Aseel M. Hamzah¹ 🝺, Tamara N. Dawood² 🕩

¹Zoonotic Disease Unit, Veterinary Medicine College, Baghdad University, Baghdad, Iraq ²Department of Public Health, College of Veterinary Medicine, Baghdad University, Baghdad, Iraq

ABSTRACT

Objective: The goal of this research was to evaluate where selenium nanoparticles impact the activity of antibodies in immunized lambs with foot and mouth vaccines by modulating the immune system.

Materials and Methods: Two groups of lambs of 3–4 months of age were injected with 1 ml of ARRIAH-VAC vaccine intramuscularly in the neck, five Lambs were given selenium nanoparticles (size 100 nm) oral administration of selenium nano dose of 0.1 mg/kg of body mass once every day for sixty days considered as group one (G1) while the other five used as control Group 2 (G2). **Results**: This resulted in the establishment of an immune response, as evidenced by a rise in antibody titer in the blood using the ELISA test for three serotypes A, O, and Asia 1, when selenium nanoparticles were given orally at a dose of 0.1 mg/kg body weight after immunization, we noticed a significant (p > 0.05) selenium nano group increase in IgG response in all immunized groups in contrast to lambs that had only received the foot-and-mouth disease vaccine

Conclusion: We have demonstrated that selenium nanoparticles administered orally significantly enhance immune responses while also increasing body weight.

Introduction

As a widespread viral animal disease that kills young animals and reduces adult animals' productivity, foot-andmouth disease (FMD) continues to be regarded for its negative social and economic effects as well as its financial costs [1]. The first confirmed FMD cases were recorded in 1937, although serotype (A) of FMD disease was not discovered until 1952 in Iraq. Since then, a number of serotypes have been identified; the serotypes O, SAT-1, and Asia 1 were identified in 1957, 1962, and 1975, respectively, and various studies from 1999 found that the most prevalent serotypes found in Iraq between 1952 and 1998 were serotype A, O, Sat1, and Asia 1A/ASIA/G-VII [G-18] is the name of the new serotype A lineage [2–5]. Nonetheless, Iraq suffers from an annual outbreak of FMD that costs livestock owners a lot of money. The disease affects cattle, buffalo, and small ruminants, with the number of sick and dead animals changing, the sickness reappeared in 2011 and affected 13,305 small ruminants, 6,757 cattle, and 5,216 buffalo. The veterinary clinic initiated a vaccination program in 2011, and about 1,935,510 buffalo, cattle, and 7,062,003 small ruminants, in that order, got immunizations, despite the fact that fewer animals were impacted overall. While the number of diseased animals was lower in 2012 when FMD outbreaks returned to fifteen Iraqi governorates, the veterinary service also launched a vaccination campaign that year, immunizing over 7,105,941 and 1,798,074 bovines and ovines, respectively. The number of animals infected with disease was noticeably declining in 2013, 2014, and 2015 while the prevalence rate for bovine and ovine increased in 2016 compared to 2015, according to the veterinary service [6].

The immunomodulatory effects of selenium nanoparticles were assessed in this work, producing long-lasting humoral and cellular immunological responses to boost vaccination-induced immune response [7–9]. Nanoselenium is

VOL 11, NO. 2, PAGES 367-375

June 2024



Received December 03, 2023 Revised January 03, 2024 Accepted February 17, 2024 Published June 08, 2024

KEYWORDS

Selenium nanoparticles; FMD vaccine; ELISA



© The authors. This is an Open Access article distributed under the terms of the Creative Commons Attribution 4.0 License (http://creativecommons.org/ licenses/by/4.0)

Correspondence: Aseel M. Hamzah 🖾 aseelm30@covm.uobaghdad.edu.iq 🗔 Zoonotic Disease Unit, Veterinary Medicine College, Baghdad University, Baghdad, Iraq.

How to cite this article: Hamzah AM, Dawood TN. Selenium nanoparticles effect on foot and mouth disease vaccine in local Awassi breed male lambs. J Adv Vet Anim Res 2024; 11(2):367–375.

less toxic as well as more suitable for biological use than sodium selenite, and it also has a number of helpful features, including excellent antioxidant activity, adsorption strength, surface activity, and catalytic performance [10-13]. Antioxidants selenium (Se) provide cell protection via reducing free radicals and preventing lipid peroxidation, from reactive oxygen (RO) [14,15] as well as the activity of glutathione peroxidases depends on selenium (Se) which enzymes are capable of reducing lipid hydroperoxides and hydrogen peroxide [16]. Additionally, Selenium serves an important role in a number of substances that alter the production of specific cytokines or improve immunological cells that are resistant to oxidative damage, both of which are known to enhance immune system response [17,18]. The amount of selenium that an animal needs relies on the minerals that are present in the soil, as well as the qualities of the food that is grown and fed to animals. The reaction to neonatal survival and juvenile animal production efficiency is contingent upon the concentration of selenium inside the animal body [19]. A supplement substitute that can provide benefits that enable needs to be satisfied by constant element release is the use of ruminal bolus containing prolonged-release nanoparticles [20]. Furthermore, Nanoselenium looks to be less toxic and more biocompatible than sodium selenite, in addition to having several beneficial qualities such as high antioxidant activity, adsorption strength, surface activity, and catalytic performance [21,22]; additionally, humoral immunity is also increasing [23,24].

Current research is therefore focused on the application of selenium nanoparticles to target enhancing the local male lambs' immune response against FMD vaccine. In particular, the experiment was designed to test whether or not selenium nanoparticles alter immunity in FMD antibody-bearing lambs that had just been injected with the vaccine. This indicates that when selenium nanoparticles were given orally following immunization, there was an increase of IgG in all the groups compared to lambs which only received FMD vaccine. This study shows that selenium nanoparticles can be a useful way to boost immune responses in vaccinated lambs.

The objective of the current research was to determine the bioavailability of selenium (Se) in lambs following its administration in the form of Se nanoparticles after vaccinating the lamb with the FMD vaccine. The goal of the current work is to evaluate the new biosynthesis of Se-nanoparticles to increase body weight and potent the immune system.

Material and Methods

Ethical statement

College of Veterinary Medicine's Institutional Animal Care and Use Committee approved every treatment performed on the study's animals (Reference number: P.G. 1301).

Bacterial culture

In 250 ml of nutrient broth, a loopful of *P. aeruginosa* was injected, where it was then left to incubate for 24 h at 37°C. Centrifugation was utilized to eliminate bacterial cells at 6,000 rpm for 10 min, after which liquid was gathered and placed in a separate 250-ml container.

Selenium nanoparticle synthesis

Bacterial cultivation in a 250 ml flask that had been previously prepared as described within the "Bacteria" portion, which includes simply Bacteria and media, was mixed with Na_2SeO_3 at a concentration of 2 mM for the nanoparticle formation. The bacteria were incubated for 72 h at 37°C after being injected with the metallic salt and 200 rpm in a shaking incubator. Within three days of the experiment, NB media's typical yellowish color changed to orange-red, showing that Se had been reduced to elemental selenium then overall samples were centrifugation at 6,000 rpm for fifteen minutes after 72 h after that the particle was washed twice with deionized distal water after the supernatant was removed then the sediment put in glasses petri dish and incubated at 37°C for dryness.

UV-visible spectra analysis

The generated spectrum of selenium nanoparticles was assessed using a JASCO 670-UV-Vis spectrophotometer with a resolution of 0.86 nm and a wavelength range of 200 to 1,200 nm at ambient temperature [25,26].

Fourier transform infrared (FTIR)

The produced selenium nanoparticles were examined by FTIR spectroscopy as explained in [27].

X-ray diffraction (XRD)

XRD (Shemadzu-6000 Japan) was used to characterize the crystalline structure of nanoparticles [28,29].

The experimental animal

The study was conducted at the Field College of Veterinary Medicine/University of Baghdad. We employed ten healthy males (local Awassi lambs) that were 3 months old and weighed, on average, between 18.5 and 20 kg. In contrast to the first group (G1), taken once a day by mouth at a dosage of 0.1 mg/kg of selenium nanoparticles for eight weeks Table 1. The composition of the elements in a concentration diet.

"Nutritional ingredients"	%
Wheat bran	20
Corn	20
Barley	48
Soya bean	10
Premix	1
Salt	1
% Total	100

while group 2 (G2) is considered as the control group. The animals were kept in the field of the veterinary medicine college and were given access to water and a daily feeding of concentrated nutrition that represents two percent and half of each body mass per head (Table 1) with straw and hay.

Blood samples

From the beginning until the end of the project, as well as zero-time blood samples have been received biweekly. Samples of blood were obtained from the jugular vein using single-use, sterile syringes after disinfecting the region where the blood was obtained. Two sets of samples were created; the first group was stored in 5 ml tubes with the anticoagulant ethyl diamine tetra acetic acid (EDTA) to estimate the white blood cell count and differential white blood cells (WBCs). The remaining blood sample was kept in sterile (10 ml) vacuum tubes. The blood sample was centrifuged for fifteen minutes at 3,000 rpm to extract the serum and measure immunological parameters and globulin content.

White blood cell count

There are five different kinds of white blood cells were distinguished and measured: neutrophils, monocytes, eosinophils, lymphocytes, and basophils). Furthermore, one drop of blood was applied to the slide and spread out, and the sample was allowed to dry for one minute before being fixed with methyl alcohol to determine the proportion of white blood cells.

The samples were then stained for three to four minutes using (Giemsa stain). After that, three drops of distilled water were added, and they were left on for two minutes. Finally, the samples were washed with tap water and dried. Following that, these were inspected using a light microscope with an oily emersion lens to identify 100 cells, and their percentages were computed as an illustration of [30].

Enzyme-linked immunosorbent assay (ELISA) test

We used a commercial ELISA kit (IDEXX[®]) to assess the IgG level, and we simultaneously examined all sera (sera group 1 and 2) with the FMDV ELISA kit.

Statistical analysis

Using SAS, data statistical analysis was carried out (System for Statistical Analysis—9.1 version). One-way analysis of variance and the least significant differences (LSDs) test were used to assess the significance of mean differences. The indicator of statistical significance is shown by (p > 0.05) [31].

RESULTS

Description of nanoparticles

With a 190–800 NM Ultra Violet—vis spectrophotometer, the absorbance properties of the isolated selenium nanoparticles were obtained (Fig. 1). The highest absorption, in particular, was observed at about 290 nm.

Pseudomonas aeruginosa bacteria

Selenium nano-particles crystallite structure produced by the biological process was analyzed using the XRD spectrum technique. A common technique for estimating the size and dimensional parameters of crystals is the use of X-rays. The Scherrer equation, which is employed for this purpose, is as follows:

- G.S= $0.9\lambda/\beta \cos\theta$
- G.S is the grain size,

B is the full width at a half maximum

 θ is the diffraction angle

 λ is the wavelength for the X-ray source used (1.5406 Å).

In FTIR, the four solutions tested: *P. aeruginosa* culture, supernatant solution after centrifugation of the nutrient cultured with *P. aeruginosa* selenium salt solution (NaHSeO₃), and the solution of selenium nanoparticles was examined using the chromatographic images obtained during the FTIR test. The results are displayed in (Fig. 3) the examination of IR absorption vibrations based on wave number (cm) allowed for the identification of many zones.

In body weight during the experiment's 60 days, the group A body weight significantly increased as shown in Table 2.

Assessment of antibody reactions

Total IgG antibodies in serum samples were quantified using the ELISA enzyme-linked immunosorbent assay.

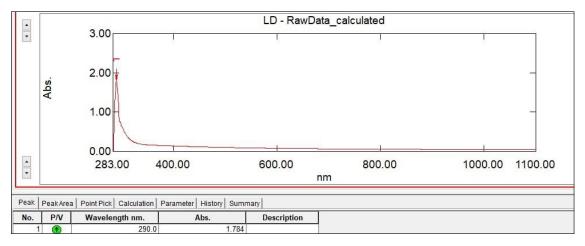


Figure 1. Absorption spectra of selenium nanoparticles isolated from Pseudomonas.

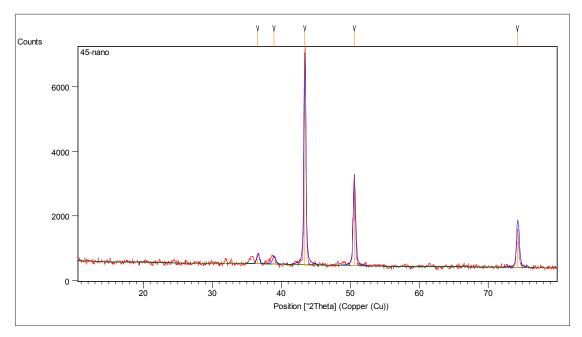


Figure 2. Displays XRD patterns for the Se nanoparticles produced using P. aeruginosa.

14, 30, 45, and 60 days following vaccination, serum samples were taken. On day 14, the antibody level was at its maximum. Following vaccination, we observed a statistically significant increase in IgG response in all immunized groups G1 when selenium nanoparticles were given orally at a concentration of 0.1 mg per kilogram of total body weight in comparison to lambs who had only received the FMD vaccine as shown in Figure 4 for three serotypes compare with control group G2.

In the present work, we suggest a possible strategy for using selenium nanoparticles to boost lambs' immunity. These results can be a consequence of selenium nanoparticles' greater absorption. Our findings imply that feeding lambs a diet enriched with selenium nanoparticles as shown in Table 3.

Discussion

The synthesis and surface plasmon vibration of selenium nanoparticles caused the UV spectra to focus in the range of two-three hundred nm [32,33]; moreover, the resonance of surface plasmon of the selenium nanoparticles made by *P. aeruginosa* is represented by the peak with the best definition at 290 nm however the present finding is consistent with findings from studies using other types of bacteria

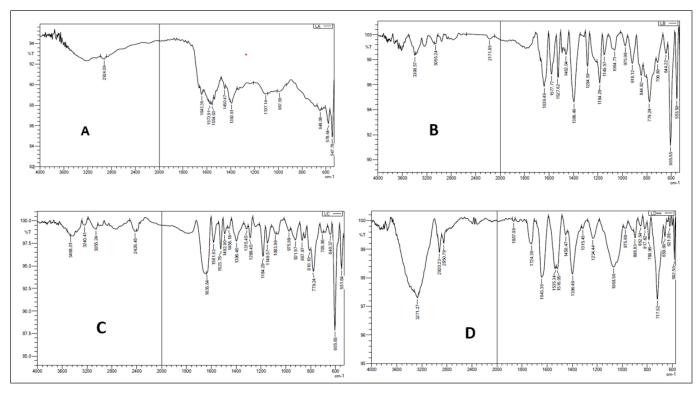


Figure 3. Illustrate the examination of the chromatographic images obtained during the FTIR test for the four solutions that were put to the test: (A) *P. aeruginosa* culture, (B) supernatant solution after centrifugation the nutrient cultured with *P. aeruginosa*, (C) selenium salt solution (NaHSeO3), and (D) selenium nanoparticle solution. Depending on the wave number (cm), different zones were detected in the vibrations induced by different infrared (IR) absorption.

Selenium nanoparticles G1	Control G2	LSD
E18.97 ± 2.14 ^a	B18.71 ± 0.88ª	
DE21.18 ± 1.82°	B20.02 ± 1.09 ^a	
CDE23.82 ± 1.78 ^a	B20.94 ± 0.94 ^a	5.57
CD26.30 ± 1.58 ^a	AB23.26 ± 0.32ª	
AB32.35 ± 1.70 ^a	AB26.44 ± 0.67 ^b	
	E18.97 ± 2.14 ^a DE21.18 ± 1.82 ^a CDE23.82 ± 1.78 ^a CD26.30 ± 1.58 ^a	E18.97 \pm 2.14°B18.71 \pm 0.88°DE21.18 \pm 1.82°B20.02 \pm 1.09°CDE23.82 \pm 1.78°B20.94 \pm 0.94°CD26.30 \pm 1.58°AB23.26 \pm 0.32°

Table 2. The effect of Se-nanoparticles on body weight during 60 days.

Means of different small letters in the same raw are significantly different (p < 0.05).. Means of different capital letters in the same column are significantly different (p < 0.05).

and fungi [34,35]. Additionally, further evidence that the artificial particles were polydisperse came from the peak's broadening [36].

The findings of this study show that selenium is the primary component of the reduction product in XRD [37] as well as the signal of the elements P, O, and C may be obtained through the extraction of a reducing agent [38].

In the infrared spectra of nanoparticles, the main absorbance bands were at 3271.27, 2920.23, 2850.79, 1867.09, 1724.36, 1643.35, 1535.34, 1516.05, 1450.47, 315.45,

1234.44, 1068.56, 975.98, 898.83, 852.54, 817.82, 786.96, 717.52, 659.66, and 621.08. The vibrations of O–H groups stretched could be responsible for the wide band observed at 3271.27/cm in addition, the vibrations that stretch asymmetrically of amide are what make up the band at 1535.34 and 1516.05 cm/1. However, the 1068.56 cm-1 peaks provide evidence that C–O exists. The sharp band may be caused by carbonyl stretching vibrations in aldehydes, ketones, and carboxylic acids at 717.52 cm⁻¹ [35,39]. Many bacterial species probably employ a range of reduction techniques, including selenite reductase

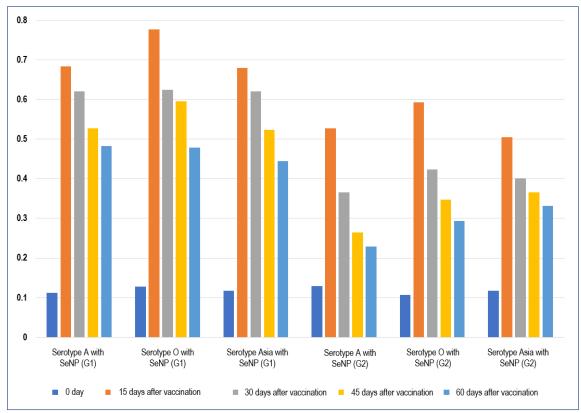


Figure 4. ELISA test for three serotypes of FMD vaccine with administration of selenium nanoparticles in group1 compare with the control group.

enzymes, membrane-associated reductase, periplasmic dissimilatory nitrite reductase, and extracellular reductases (nitrate reductase) [40,41]. These findings showed that these bioactive compounds' functional groups may potentially operate when synthesizing selenium nanoparticles, as reduction and stabilization agents.

The analytical technique of XRD is primarily used to identify crystalline nano materials and can provide details on the size of the nano material and the type of crystal in the compound. Angle Theta-2 value is used in analytical XRD for varied peaks [42].

The prepared selenium nanoparticles demonstrated increased body weight mass gain. The process that selenium might cause a rise in excess weight may be linked to changes in hepatic fatty acid metabolism and energy production by raising the Glut2 transcription in addition to the lipid metabolism-related enzymes [43].

Selenium nanoparticles have been shown to interact with globulin proteins through direct binding interactions, modulation of protein structure and function, enhancement of antioxidant activity, and regulation of gene expression. These interactions suggest a correlation between selenium nanoparticles and globulin, highlighting the potential impact of selenium nanoparticles on immune function and overall health [44,45].

The impact of selenium supplementation or depletion on cellular and humoral immunological reply to various vaccinations has been well studied in studies employing agricultural animals [46]. It is thought that Se's incorporation into selenium-proteins is responsible for its immunomodulatory actions. Among selenium-proteins, selenoenzymes such as thioredoxin reductases and glutathione peroxidases control the proportion of redox signals to RO species by eliminating too many potentially harmful radicals generated when oxidative damage occurs in immune cells [47,48].

The host's immunological response against FMD infection depends significantly on humoral immunity. Sandwich ELISA examination of blood samples revealed that, in comparison to a control group, oral administration of selenium nanoparticles followed by injection of the FMD vaccination has significantly improved antibody responses (p < 0.05); however, the increased immunological responses found in this study are consistent with earlier studies [49].

Table 3. Correlation coefficient between different deferentialWBCs after vaccination with the FMD vaccine.

Parameters	Correlation coefficient-r			
	ELISA type A	ELISA type O	ELISA type Asia 1	
Lymphocyte	0.783**	0.8602**	0.8674**	
Neutrophil	0.7871**	0.9116**	0.8278**	
Monocyte	0.35**	0.34**	0.50**	
Basophil	0.10 NS	0.11 NS	0.11 NS	
Eosinophil	0.02 NS	-0.15 NS	-0.02 NS	
Globulin	0.8993**	0.8813**	0.872**	

**(p<0.01), NS: Non-Significant.

Conclusion

In this paper, we provide a possible strategy to enhance lamb immunity through the use of selenium nanoparticles. The increased selenium nanoparticle absorption may be the cause of these findings. Our findings indicate that providing lambs with a meal supplemented with selenium nano could potentially stimulate their immune system. Selenium nanoparticles, on the other hand, may facilitate nutritional digestion; accessible selenium, on the other hand, could raise protein creation via enhancing stimulation of many cellular metabolic processes, leading to improved mRNA expression for protein synthesis. As well as food intake and weight gain were both boosted by taking supplements of selenium nanoparticles.

List of Abbreviations

cm: centimeter; EDTA: ethyl diamine tetra acetic acid; ELISA: enzyme linked immunosorbent assay; FMD: foot and mouth disease; FTIR: Fourier transform infrared spectroscopy; G1: group one; G2: group two; IgG: immunoglobulin type G; IR: Infrared; LSD: least significant differences; NM: nanometer; NMD: nutritional muscular dystrophy; RO: reactive oxygen; Se: selenium; UV: ultra violet; WBCs: white blood cells; XRD: X-ray diffraction analysis.

Acknowledgment

Special appreciation to the College of Veterinary Medicine for providing and supporting this work, special thanks, and gratitude goes out to the staff at the Virology Department of Veterinary Laboratory and research-veterinary director else gratitude goes to all the professors and personnel of the Department of Veterinary Public Health.

Conflict of interest

In relation to the experiments and the publication, the authors declared no conflicts of interest.

Authors' contributions

TND supervised the research project; AMH conducted the experiments and produced the original paper. AMH and TND also developed and designed the experiments and edited and evaluated the text. All authors revised and approved the final manuscript.

References

- Jamal SM, Belsham GJ. Foot-and-mouth disease: past, present and future. Vet Res 2013; 44(1):1–14. Available via http://www.veterinaryresearch.org/content/44/1/116
- [2] Mahdi AJ. Foot and mouth disease in Iraq: strategy and control. Master of Science Department of Diagnostic Medicine/ Pathobiology College of Veterinary Medicine Kansas State University, Manhattan, Kansas, 2010. Available via http://hdl.handle.net/2097/4620
- [3] Hussein ZS. foot and mouth disease in some areas of Nainawa province during may 2007. Iraqi J Vet Med 2008; 32(2):181– 203. Available via https://www.iasj.net/iasj/download/ bf81a210a96aff33
- [4] Sabar AA, Al-Banna AS, Abdul-Rasoul LMS, Abood BK. Diagnostic study of FMD virus in different area in Iraq. Iraqi J Vet Med 2012; 36(0E): 49–55. Available via https://www.iasj.net/iasj/ download/0895af282bc4077e
- [5] Jumaa RS, Mohsin SI, Abdulmjeed DI, Atshan OF. Foot and mouth disease virus: a review. Magna Sci Adv Biol Pharm 2021; 03(02):027– 035; https://doi.org/10.30574/msabp.2021.3.2.0038
- [6] Al-Salihi KA. The epidemiology of foot-and-mouth disease outbreaks and its history in Iraq. Vet World 2019; 12(5):706; https:// doi.org/10.14202/vetworld.2019.706-712
- [7] Bagheri-Josheghani S, Bakhshi B. Formulation of selenium nanoparticles encapsulated by alginate-chitosan for controlled delivery of Vibrio Cholerae LPS: a novel delivery system candidate for nanovaccine. Inter J Biol Macro 2022; 208:494–508; https:// doi.org/10.1016/j.ijbiomac.2022.03.087
- [8] Khabatova VV, Serov DA, Tikhonova IV, Astashev ME, Nagaev EI, Sarimov RM, et al. Selenium nanoparticles can influence the immune response due to interactions with antibodies and modulation of the physiological state of granulocytes. Pharma 2022; 14(12):2772;https://doi.org/10.3390/ pharmaceutics14122772
- [9] Mal'tseva VN, Gudkov SV, Turovsky EA. Modulation of the functional state of mouse neutrophils by selenium nanoparticles *in vivo*. Inter J Mol Sci 2022; 23(21):13651; https://doi.org/10.3390/ ijms232113651.
- [10] Zhang JS, Wang XF, Xu TW. Elemental selenium at nano size (nano-Se) as a potential chemo preventive agent with reduced risk of selenium toxicity: comparison with Se-methyl selenocysteine in mice. Toxico Sci 2008; 101:22–31; https://doi.org/10.1093/ toxsci/kfm221
- [11] Boostani A, Sadeghi AA, Mousavi SN, Chamani M, Kashan N. Effects of organic, inorganic, and nano-Se on growth performance, antioxidant capacity, cellular and humoral immune responses in broiler chickens exposed to oxidative stress. Live Sci 2015; 178:330–6; https://doi.org/10.1016/j. livsci.2015.05.004

- [12] Skalickova S, Milosavljevic V, Cihalova K, Horky P, Richtera L, Adam V. Selenium nanoparticles as a nutritional supplement. Nutr 2017; 33: 83–90; https://doi.org/10.1016/j.nut.2016.05.001
- [13] Wali AT, Alqayim MAJ. Biosynthesis, characterization and bioactivity of selenium nanoparticles synthesized by propolis. Iraqi J Vet Med 2019; 43(1):197–209; https://www.iasj.net/iasj/ article/167214
- [14] Harsini SG, Habibiyan M, Moeini MM, Abdolmohammadi AR. Effects of dietary selenium, vitamin E, and their combination on growth, serum metabolites, and antioxidant defense system in skeletal muscle of broilers under heat stress. Biol Trace Elem Res 2012; 148 (3):322–330; https://doi.org/10.1007/s12011-012-9374-0
- [15] Akbari B, Baghaei-Yazdi N, Bahmaie M, Mahdavi Abhari F. The role of plant-derived natural antioxidants in reduction of oxidative stress. BioFactors 2022; 48(3):611–33; https://doi.org/10.1002/ biof.1831
- [16] Shen Y, Huang H, Wang Y, Yang R, Ke X. Antioxidant effects of Se-glutathione peroxidase in alcoholic liver disease. J Trace Ele Med Biol 2022; 74:127048; https://doi.org/10.1016/j. jtemb.2022.127048
- [17] De GA, Riva L, Trujillo LA, González-Hernández JC. Assessment on oxidative stress in animals: from experimental models to animal production. In: importance of oxidative stress and antioxidant system in health and disease. IntechOpen, London, UK, 2022; https:// doi.org/10.5772/intechopen.109043
- [18] Li Y, Fan M, Qiu Q, Wang Y, Shen X, Zhao, K. Nano-selenium and Macleaya cordata extracts improved immune function and reduced oxidative damage of sows and IUGR piglets after heat stress of sows in late gestation. Biol Trace Ele Res 2022; 200(12):5081–90; https://doi.org/10.1007/s12011-022-03103-y
- [19] Saha U, Fayiga A, Hancock D, Sonon, L. Selenium in animal nutrition: deficiencies in soils and forages, requirements, supplementation and toxicity. Inter J Appl Agri Sci 2016; 2(6):112–25; https:// doi.org/10.11648/j.ijaas.20160206.15
- [20] Bribiesca JER, Casas RL, Monterrosa RGC, Pérez AR. Supplementing selenium and zinc nanoparticles in ruminants for improving their bioavailability meat. In Nutrient delivery. Academic Press, Cambridge, MA, pp 713–47, 2017; https://doi.org/10.1016/ B978-0-12-804304-2.00019-6
- [21] HashemAH,AbdelazizAM,AttiaMS,SalemSS.Seleniumandnano-selenium-mediated biotic stress tolerance in plants. In Selenium and nano-selenium in environmental stress management and crop quality improvement. Springer, Cham, pp 209–26, 2022; https:// link.springer.com/chapter/10.1007/978-3-031-07063-1_11
- [22] Chen N, Yao P, Zhang W, Zhang Y, Xin N, Wei H, et al. Selenium nanoparticles: enhanced nutrition and beyond. Crit Rev Food Sci Nut 2023; 63(33):12360–71; https://doi.org/10.1080/10408398 .2022.2101093
- [23] Bami MK, Afsharmanesh M, Espahbodi M, Esmaeilzadeh E. Effects of dietary nano-selenium supplementation on broiler chicken performance, meat selenium content, intestinal microflora, intestinal morphology, and immune response. J Trace Ele Med Biol 2022; 69:126897; https://doi.org/10.1016/j.jtemb.2021.126897
- [24] Liu C, Li Y, Li H, Wang Y, Zhao K. Nano-selenium and Macleaya cordata extracts improved immune functions of intrauterine growth retardation piglets under maternal oxidation stress. Biol Trace Elem Res 2022; 200(9):3975–82; https://doi.org/10.1016/j. jtemb.2021.126897
- [25] Mazhir SN, Abdalameer NK, Yaaqoob LA, Hammood JK. Biosynthesis of (Zn/Se) core-shell nanoparticles by micro plasma-jet technique. Inter J Nano 2022; 21(5):2250041–914; https://doi. org/10.1142/S0219581X22500417
- [26] Mohammed S, Ali AH, Adem K, Yaaqoob L. Antibacterial activity with zeolitic nano-particles activated by microwave plasma. Egyp J Chem 2022; 65(2):161–6; https://doi.org/10.21608/ ejchem.2021.83414.4091

- [27] Augustine SK, Bhavsar SP, Kapadnis BP. A non-polyene antifungal antibiotic from Streptomyces albidoflavus PU 23. J Biosci 2005; 30(2):201–11; https://link.springer.com/article/10.1007/ BF02703700
- [28] Abdalameer NK, Mazhir SN, Aadim KA. The effect of ZnSe Core/ shell on the properties of the window layer of the solar cell and its applications in solar energy. Ener Rep 2020; 6:447–58; https:// doi.org/10.1016/j.egyr.2020.09.023
- [29] Mazhir SN, Abdalameer NK, Yaaqoob LA, Hammood JK. Cold plasma synthesis of Zinc selenide nanoparticles for inhibition bacteria using disc diffusion. Phys Chem Solid State 2022; 23(4):652– 8; https://doi.org/10.15330/pcss.23.4.652-658
- [30] Oktiyani N, Muhlisin A, Roebiakto E, Norsiah W, Mahpolah, M. Utilization of alternative buffer solutions for staining thin blood smears by the giemsa, wright stain and romanowsky method. Trop Health Med Res 2023; 5(1):34–45; https://doi.org/10.35916/ thmr.v4i1.76
- [31] SAS. Statistical Analysis System, User's Guide. Statistical.; Version 9.6th ed., SAS Institute Inc., Cary, NC, 2018.
- [32] Yang LB, Shen YH, Xie AJ, Liang JJ, Zhang BC. Synthesis of Se nanoparticles by using TSA ion and its photocatalytic application for decolorization of congo red under UV irradiation. Mat Res Bull 2008; 43:572–82; https://doi.org/10.1016/j.materresbull.2007.04.012
- [33] Mishra RR, Prajapati S, Das J, Dangar TK, Das N, Thatoi H. Reduction of selenite to red elemental selenium by moderately halotolerant Bacillus megaterium strains isolated from Bhitarkanika mangrove soil and characterization of reduced product. Chemosphere 2011; 84:1231–7; https://doi.org/10.1016/j.chemosphere.2011.05.025
- [34] Fesharaki PJ, Nazari P, Shakibaie M, Rezaie S, Banoee M, Abdollahi M, et al. Biosynthesis of selenium nanoparticles using Klebsiella pneumoniae and their recovery by a simple sterilization process. Braz J Microbio 2010; 41:461–6; https://doi.org/10.1590/ S1517-83822010000200028
- [35] Zhang W, Chen Z, Liu H, Zhang L, Gao P, Li D. Biosynthesis and structural characteristics of selenium nanoparticles by Pseudomonas alcaliphila. Coll Sur B Bio 2011; 88(1):196–202; https://doi. org/10.1016/j.colsurfb.2011.06.031
- [36] Reddy V, Torati RS, Oh S, Kim C. Biosynthesis of gold nanoparticles assisted by Sapindus mukorossi Gaertn. Fruit Pericarp and their catalytic application for the reduction of p-Nitroaniline. Indus Eng Chem Res 2013; 52:556–64; https://doi.org/10.1021/ie302037c
- [37] Shakibaie M, Khorramizadeh M, Faramarzi M, Sabzevari O, Shahverdi A. Biosynthesis and recovery of selenium nanoparticles and the effects on matrix metalloproteinase-2 expression. Biotech Appl Bioch 2010; 56(1):7; https://doi.org/10.1042/BA20100042
- [38] Dhanjal S, Cameotra SS. Aerobic biogenesis of selenium nanospheres by Bacillus cereus isolated from coalmine soil. Micro Cell fact 2010; 9(1):1–11; https://doi.org/10.1186/1475-2859-9-52
- [39] Kora AJ, Rastogi L. Biomimetic synthesis of selenium nanoparticles by *Pseudomonas aeruginosa* ATCC 27853: an approach for conversion of selenite. J Environ Manage 2016; 181:231–6; https://doi. org/10.1016/j.jenvman.2016.06.029
- [40] Lampis S, Zonaro E, Bertolini C, Bernardi P, Butler CS, Vallini G. Delayed formation of zero-valent selenium nanoparticles by Bacillus mycoides SeITE01 as a consequence of selenite reduction under aerobic conditions. Micro Cell fact 2014; 13(1):1–14; https://doi.org/10.1186/1475-2859-13-35
- [41] Dwivedi S, AlKhedhairy AA, Ahamed M, Musarrat J. Biomimetic synthesis of selenium nanospheres by bacterial strain JS-11 and its role as a biosensor for nanotoxicity assessment: a novel Se-bioassay. PloS One 2013; 8(3):e57404; https://doi. org/10.1371/journal.pone.0057404
- [42] Chandramohan S, Sundar K, Muthukumaran, A. Hollow selenium nanoparticles from potato extract and investigation of its biological properties and developmental toxicity in zebrafish embryos.

IET Nano 2019; 13(3):275-81; https://doi.org/10.1049/ iet-nbt.2018.5228

- [43] Hu X, Chandler JD, Orr ML, Hao L, Liu K, Uppal K, et al. Selenium supplementation alters hepatic energy and fatty acid metabolism in mice. J Nut 2018; 148(5):675–84; https://doi.org/10.1093/jn/ nxy036
- [44] Ferreira RLU, Sena-Evangelista KCM, De Azevedo EP, Pinheiro FI, Cobucci RN, Pedrosa, LFC. Selenium in human health and gut microflora: bioavailability of selenocompounds and relationship with diseases. Fron Nut 2021; 8:685317; https://doi.org/10.3389/ fnut.2021.685317
- [45] Jampilek J, Kralova K. Potential of nanonutraceuticals in increasing immunity. Nano 2020; 10(11):2224; https://doi.org/10.3390/ nano10112224
- [46] Spallholz JE, Boylan LM, Larsen HS. Advances in understanding selenium's role in the immune system. Anna New York Acad Scien

1990; 587:123–39. Available via http://acikerisim.akdeniz.edu.tr/xmlui/handle/123456789/3668

- [47] Hoffmann PR. Mechanisms by which selenium influences immune responses. Arch Immunol Therap Exper 2007; 55:289; https://doi. org/10.1007/s00005-007-0036-4
- [48] Huang Z, Rose AH, Hoffmann PR. The role of selenium in inflammation and immunity: from molecular mechanisms to therapeutic opportunities. Antiox Red Sign 2012; 16:705–43; https://doi. org/10.1089/ars.2011.4145
- [49] Maqbool B, Wang Y, Cui X, He S, Guan R, Wang S, et al. Ginseng stem-leaf saponins in combination with selenium enhance immune responses to an attenuated pseudorabies virus vaccine. Microbiol Immuno 2019; 63(7):269–79; https://doi. org/10.1111/1348-0421.12715