SHORT COMMUNICATION

Isolation of multi-drug resistant *Klebsiella* sp. from bovine mastitis samples in Rangpur, Bangladesh

Md. Salauddin, Mir Rowshan Akter, Md. Khaled Hossain, Md. Mostafizer Rahman

Department of Microbiology, Faculty of Veterinary and Animal Science, Hajee Mohammad Danesh Science and Technology University, Dinajpur, Bangladesh

ABSTRACT

Objective: The objective of this study was to identify the multi-drug resistance (MDR) *Klebsiella* sp. from mastitis milk samples.

Materials and Methods: In the current research, 48 clinical mastitis milk samples were collected from Rangpur division, Bangladesh. Confirmation of bovine mastitis (BM) was done by the California Mastitis Test (CMT). All the CMT positive isolates were subjected for the identification of *Klebsiella* sp. using through a series of cultural and biochemical tests. MDR *Klebsiella* sp. isolates were determined using the disk diffusion method, and minimum inhibitory zones were measured by following Clinical and Laboratory Standards Institute. MDR patterns of the isolates were also subjected to study by using housefly (*Musca domestica*).

Results: Among the isolates, 62.5% (*n* = 30/48) revealed the presence of *Klebsiella* sp. Eight antimicrobial agents including Amoxicillin, Novobiocin, Erythromycin, Vancomycin, Cephradine, Tetracycline, Bacitracin, Methicillin, and housefly (*M. domestica*) showed complete resistance to *Klebsiella* sp. On the other hand, Chloramphenicol, Gentamicin, Ciprofloxacin, Azithromycin, Norfloxacin, Levofloxacin, and Nalidixic acid showed sensitivity.

Conclusion: This study helps to treat BM with effective antibiotics and helps in an epidemiological study in Rangpur division as well as helps to create public health awareness.

ARTICLE HISTORY

Received April 15, 2019 Revised April 30, 2019 Accepted May 14, 2019 Published July 25, 2019

KEYWORDS

Antibiogram; cattle; *Klebsiella* sp.; mastitis



This is an Open Access article distributed under the terms of the Creative Commons Attribution 4.0 Licence (http://creativecommons.org/ licenses/by/4.0)

Introduction

Antimicrobial resistance is an ultimate threat to the animal as well as a human being throughout the world. Bovine mastitis (BM) is caused by a variety of bacteria; among them, *Klebsiella* sp. is an important Gram-negative pathogen which may initiate emerging incidence [1,2]. *Klebsiella* sp. is an opportunistic bacterium that can cause primary bacteremia as well as urinary tract infection in human and animal [3,4]. Fey et al. [5] reported that *Klebsiella* sp. has zoonotic importance. *Klebsiella* sp. is notoriously appeared in dairy food products [6], and it is reported that they are responsible for clinical as well as subclinical BM [7]. It is quite difficult to control BM originated from *Klebsiella* sp. infection [4]. As reported by Grohn et al. [8], milk production falls and mortality increased in cows affected with *Klebsiella* sp. They are able to produce a significant loss in the dairy farm by reducing production; which is considered as more fatal as compared to infection caused by *Escherichia coli* [9].

Extensive use of antibiotic leads to the development of multi-drug resistance (MDR) organisms. The rate of MDR organism development is increasing day by day [11]; the development of MDR *Klebsiella* sp. is also gradually increasing worldwide [11]. Consequently, both antibiotic treatment and mass vaccination showed limited effects against BM caused by *Klebsiella* sp. [12]. Increasing MDR bacteria and their treatment with antimicrobial agents as well as zoonotic importance are considered as important issues globally [13,14]. Constrained examines have been completed on the detachment of *Klebsiella* sp. in Bangladesh [15,16]. Previously, we isolated and identified *Klebsiella* sp. by conventional bacteriological techniques.



Correspondence Mir Rowshan Akter 🖾 akter.rowshan@gmail.com 🖬 Department of Microbiology, Faculty of Veterinary and Animal Science, Hajee Mohammad Danesh Science and Technology University, Dinajpur, Bangladesh.

How to cite: Salauddin M, Akter MR, Hossain MK, Rahman MM. Isolation of multi-drug resistant *Klebsiella* sp. from bovine mastitis samples in Rangpur, Bangladesh. J Adv Vet Anim Res 2019; 6(3):362–365.

The present study focused on molecular detection of mastitis-causing *Klebsiella* sp. from clinical mastitis milk samples in Rangpur division, Bangladesh, and the antibiotic susceptibility patterns of the organism were investigated for the first time in Bangladesh.

Materials and Methods

Collection and preparation of samples

A total of 48 milk samples were gathered from the selected BM dairy cows in Rangpur division, Bangladesh. The samples were collected based on clinical sign and inflammatory lesion of udder and teat. About 10–15 ml sample was collected from each dairy cow. Immediately after collection, the California Mastitis Test (CMT) was done according to Schalm and Noorlander [17] for the confirmation of BM. All the suspected samples were aseptically transferred to the Microbiology Laboratory, Hajee Mohammad Danesh Science and Technology University (HSTU) by maintaining a cool chain for microbiological analysis.

Isolation and identification of Klebsiella sp.

Samples were cultured on nutrient agar (NA), Eosin Methylene Blue (EMB) agar, and MacConkey (MC) agar at 37°C for 24 h. Isolation and identification were done by conventional techniques according to Edwards and Ewing [18]. Furthermore, the isolates were biochemically confirmed based on Merchant and Packer [19].

Antimicrobial susceptibility testing

Disk diffusion method [20] was used to determine the MDR Klebsiella sp. from the isolates using MHA (Hi-Media, India), and the zone of inhibition was interpreted according to standards of the National Committee for Clinical Laboratory Standards [21]. A total of 15 antibacterial disks (Hi-Media, India) were used in this study, namely, Gentamicin (GEN 10 µg), Amoxicillin (AMX 30 μg), Chloramphenicol (C 30 μg), Ciprofloxacin (CIP 5 μg), Bacitracin (B 10 μg), Azithromycin (AZM 30 μg), Erythromycin (E 15 µg), Methicillin (Met 5 µg), Novobiocin (NV 30 μg), Vancomycin (VA 30 μg), Norfloxacin (NX 10 μg), Tetracycline (TE 30 μg), Levofloxacin (LE 5 μg), Nalidixic acid (NA 30 µg), and Cephradine (CH 30 µg). The zones were estimated in millimeter and resistance and susceptibility were recorded [22]. These MDR Klebsiella sp. were also studied by using housefly on MHA media and observed their antimicrobial activity. On the other hand, Nazari et al. [23] studied with housefly maggot extracts, but here we applied the whole fly.

Results

The collected samples were inoculated on NA in which they produced large, circular, smooth, and convex colonies.

Round, pink, slightly raised, translucent, and mucoid colonies were found in MC, and on EMB they also showed mucoid pink colonies. Then Gram-negative, short rod with capsule *Klebsiella* sp. was observed under a microscope. The identified isolates were subjected to a biochemical test for more confirmation (Fig. 1). In methyl-red test and indole test, the isolates were produced a negative result. The Voges–Proskauer test, Simmon's citrate test, and catalase tests were positive for *Klebsiella* sp. On Triple Sugar Iron (TSI) test, the slant was yellowish with no changes in butt and no H2S produced, but gas bubble appeared.

Antimicrobial susceptibility test of *Klebsiella* sp. (Fig. 2) reveals that this organism was MDR of which AMX, B, E, MET, NV, VA, TE, and CH were completely resistant. Out of 15 antibiotic agents, GEN (19 mm), C (24 mm), CIP (30 mm), AZM (25 mm), NX (25 mm), LE (22 mm), and NA (17 mm) were showed above-mentioned zone of inhibition in mm. The positive *Klebsiella* sp. was studied using housefly and showed no zone of inhibition.

Discussion

In the present research work, 62.5% (*n* = 30) BM involved with *Klebsiella* sp. could be detected based on CMT, cultural, and biochemical tests. After the collection of mastitis milk, samples were transferred to the laboratory maintaining the cool chain. Then, grown into NA, EMB, and MC, respectively, by following Edwards and Ewing [18]. From the cultural and Gram staining test, Gram-negative, rod-shaped, and non-motile *Klebsiella* sp. were identified. From the pure culture, several different biochemical tests were performed for the confirmation of *Klebsiella* sp.

Klebsiella sp. was notoriously and ubiquitously appeared in milk along with their products that have zoo-notic importance [14]. In this research work, 30 samples were positive for *Klebsiella* sp. The prevalence of *Klebsiella* sp. in the current study was higher than the study of Gundogan and Yakar [24] and Haryani et al. [25]. This variation might be due to geographical distribution, biosecurity, and immunological status of the study population.

Antibiogram study revealed that all the isolates were showed MDR in which AMX, B, E, MET, NV, VA, TE, and CH were completely resistant to *Klebsiella* sp. which is supported by Gundogan et al. [11]. Then again, CIP (30 mm) produced the highest zone of inhibition and AZM, NX, LE, and NA were produced 25, 25, 22, and 17 mm zone of inhibition, respectively. In the present study, houseflies (*Musca domestica*) were caught and stored into PBS (Phosphate Buffer Saline) then directly placed on MHA (Mueller Hinton Agar) plates which were pre-stained with pure field isolates. Nazari et al. [23] worked with visceral parts of housefly maggot and its extracted material which showed good antimicrobial activity against different antibiotic agents. But, houseflies in the current research showed complete resistance during antibiogram study. This might be due

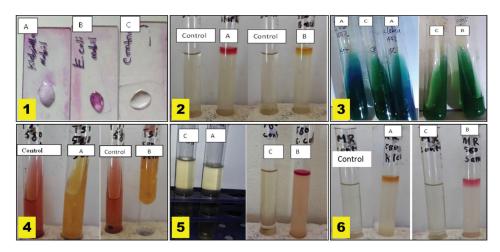


Figure 1. Biochemical tests for the identification of *Klebsiella* sp. (1) Catalase test; (A) *Klebsiella* sp., (B) *E. coli*, (C) control, (2) Voges–Proskauer test; (A) *Klebsiella* sp. positive, (B) *E. coli* negative (3) Simmon's citrate test; (A) *Klebsiella* sp. positive, (B) *E. coli* negative, (C) control (4) TSI test; (A) *Klebsiella* sp., (B) *E. coli*, (5) Indole test; (A) *Klebsiella* sp., (B) *E. coli*, (C) control, (6) Methyl-Red test; (A) *Klebsiella* sp. negative, (B) *E. coli* positive, (C) control.

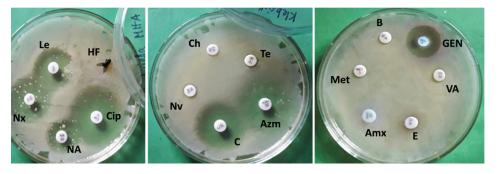


Figure 2. Antibiogram of *Klebsiella* sp. on MHA. Le = Levofloxacin, HF = Housefly, Nx = Norfloxacin, NA = Nalidixic acid, Cip = Ciprofloxacin, Ch = Cephradine, Te = Tetracycline, NV = Novobiocin, C = Chloramphenicol, Azm = Azithromycin, B = Bacitracin, Met = Methiclillin, Gen = Gentamicin, VA = Vancomycin, E = Erythromycin, Amx = Amoxicillin.

to the use of whole housefly and in external body parts of housefly carry several different organisms [26] which may be MDR. The deliberate use of antibiotic for the treatment of BM causes MDR which is a global issue. From this study, it is concluded that CIP, AZM, NX, LE, and NA can be the choice of drugs for treating the BM in Rangpur, Bangladesh.

Conclusion

The prevalence of the *Klebsiella* sp. in mastitis milk was found as 62.5%. Current research work may help to choose a specific drug to treat BM and also helps to control the indiscriminate use of antibiotics that causes MDR.

Acknowledgments

The researchers would like to thank BARC for technical support and National Institute of Biotechnology (NIB),

Savar, Dhaka to provide laboratory facility for this research and also grateful to Department of Microbiology, HSTU.

Conflict of interest

The authors declare that they have no conflict of interests.

Authors' contributions

MS, MRA, and MKH designed and interpreted experiments. MS conducted the actual experiments and prepared the draft of the manuscript. All the authors finally approved the manuscript for publication.

References

 Zadoks RN, Munoz MA. The emergence of *Klebsiella* as a major mastitis organism. Natl. Mastitis Counc. 46th Annu. Mtg., San Antonio, TX. National Mastitis Council Inc., Verona, WI, pp 100–111, 2007.

- [2] Saini V, McClure JT, Leger D, Keefe GP, Scholl DT, Morck DW, et al. Antimicrobial resistance profiles of common mastitis pathogens on Canadian dairy farms. J Dairy Sci 2012; 95:4319–32; https:// doi.org/10.3168/jds.2012-5373
- [3] Manikandan C, Amsath A. Antibiotic susceptibility pattern of *Klebsiella pneumoniae* isolated from urine samples. Int J Curr Microbiol Appl Sci 2013; 2(8):330–7.
- [4] Mansour Ahmed MA, Zaki Hoda M, Hassan Nibal A, Al-Humiany Abdulrahman A. Molecular characterization and immunoprotective activity of capsular polysaccharide of *Klebsiella Pneumoniae* isolated from farm animals at Taif Governorate. Am J Infect Dis 2014; 10(1):1–14; https://doi.org/10.3844/ajidsp.2014.1.14
- [5] Fey PD, Safranek TJ, Rupp ME, Dunne EF, Ribot E, Iwen PC, et al. Ceftriaxone-resistant salmonella infection acquired by a child from cattle. N Engl J Med 2000; 342:1242–9; https://doi.org/10.1056/ NEJM200004273421703
- [6] Sukhon SNE. Identification and characterization of *Klebsiellae* isolated from milk and milk products in Jordan. Food Microbiol 2003; 20:225–30; https://doi.org/10.1016/S0740-0020(02)00085-0
- [7] Hammad AM, Ahmed AM, Ishida Y, Shimamoto T. First characterization and emergence of SHV-60 in raw milk of a healthy cow in Japan. J Vet Med Sci 2008; 70:1269–72; https://doi.org/10.1292/ jvms.70.1269
- [8] Grohn YT, Wilson DJ, Gonzalez RN, Hertl JA, Schulte H, Bennett G, et al. Effect of pathogen-specific clinical mastitis on milk yield in dairy cows. J Dairy Sci 2004; 87:3358–74; https://doi.org/10.3168/jds. S0022-0302(04)73472-4
- [9] Langoni H, Guiduce MVS, Nobrega DB, Silva RC, Richini-Pereira VB, Salina A, et al. Research of *Klebsiella pneumoniae* in dairy herds. Pesqui Vet Brasil 2015; 35(1):9–12; https://doi.org/10.1590/ S0100-736X2015000100003
- [10] Uddin MA, Hasan M, Haque M, Noor R. Isolation and identification of pathogenic *Escherichia coli, Klebsiella* spp. and *Staphylococcus* spp. in raw milk samples collected from different areas of Dhaka city, Bangladesh. Stamford J Microbiol 2011; 1(1):19–23; https:// doi.org/10.3329/sjm.v1i1.9098
- [11] Gundogan N, Citak S, Yalcin E. Virulence properties of extended spectrum *b*-Lactamase–producing *Klebsiella* species in meat samples. J Food Prot 2011; 74(4):559–64; https://doi. org/10.4315/0362-028X.JFP-10-315
- [12] Munoz MA, Welcome FL, Schukken YH, Zadoks RN. Molecular epidemiology of two *Klebsiella pneumoniae* mastitis outbreaks on a dairy farm in New York State. J Clin Microbiol 2007; 45(12):3964– 71; https://doi.org/10.1128/JCM.00795-07
- [13] Unakal CG, Kaliwal BB. Prevalence and antibiotic susceptibility of *Staphylococcus aureus* from bovine mastitis. Vet World 2010; 3(2):65–7.
- [14] Mohanty NN, Das P, Pany SS, Sarangi LN, Ranabijuli S, Panda HK. Isolation and antibiogram of *Staphylococcus*, *Streptococcus* and

E. coli isolates from clinical and subclinical cases of bovine mastitis, Vet World 2013; 6(10):739–43; https://doi.org/10.14202/ vetworld.2013.739-743

- [15] Manu MMR. Prevalence, risk factors, and antimicrobial activity of alovera gel against the bacterial pathogens of mastitis in dairy cows at Dinajpur district of Bangladesh. MS thesis, Hajee Mohammad Danesh Science and Technology University (HSTU), Dinajpur-5200, Bangladesh, 2018.
- [16] Mia MT. Detection of bacterial spp. from clinical mastitis in dairy cows at Nilphamari District and their antibiogram study. Master's thesis, Hajee Mohammad Danesh Science and Technology University (HSTU), Dinajpur-5200, Bangladesh, 2016.
- [17] Schalm OW, Noorlander DO. Experimental and observation leading to development of California mastitis test. J Am Vet Med Assoc 1957; 130:199–204.
- [18] Edwards PR, Ewing WH. Edwards and Ewing's Identification of Enterobacteriaceae. 4th edition, Elsevier Science Publishing Co., Inc., New York, NY, 1986.
- [19] Merchant IA, Packer RA. Veterinary bacteriology and virology. 7th edition, The Iowa University Press, Ames, IA, pp 286–306, 1967.
- [20] Bauer AW, Kirby WMM, Sherris JC, Turck, M. Antibiotic sensitivity testing by a standardized single disk method. Am J Clin Path 1966; 45:493–6; https://doi.org/10.1093/ajcp/45.4_ts.493
- [21] CLSI. Performance standards for antimicrobial susceptibility testing, twenty-third informational supplement, CLSI Document M100-S23. Clinical and Laboratory Standards Institute, Wayne, PA, 2013.
- [22] Cappuccino JC, Sherman N. Microbiology. A Laboratory manual. 10th edition, United States of America, pp 293–299, 2014.
- [23] Nazari M, Mehrabi T, Hosseini SM, Alikhani MY. Bacterial contamination of adult house flies (*Musca Domestica*) and sensitivity of these bacteria to various antibiotics, captured from hamadan city, Iran. J Clin Diag Res 2017; 11(4):4–7; https://doi.org/10.7860/ JCDR/2017/23939.9720
- [24] Gundogan N, Yakar U. Siderophore production, serum resistance, hemolytic activity and extended spectrum beta lactamase- producing *Klebsiella* species isolated from milk and milk products. J Food Safety 2007; 3:251-60; https://doi. org/10.1111/j.1745-4565.2007.00077.x
- [25] Haryani Y, Noorzaleha AS, Fatimah AB, Noorjahan BA, Patrick GB, Shamsinar AT, et al. Incidence of *Klebsiella pneumoniae* in street foods sold in Malaysia and their characterization by antibiotic resistance, plasmid profiling, and RAPD-PCR analysis. Food Control 2007; 18:847–53; https://doi.org/10.1016/j. foodcont.2006.04.009
- [26] Ghalehnoo MR. Housefly (*Musca domestica*) as carrier of Enterotoxigenic *Staphylococcus aureus* in broiler farms in Iran: is it important for public health? Int J Enteric Pathog 2015; 3(3):e25688; https://doi.org/10.17795/ijep25688