**Original Article** 

**September 2017** Vol 4 No 3, Pages 295-300.



# Prevalence and identification of subclinical mastitis in cows at BLRI Regional Station, Sirajganj, Bangladesh

Md. Humayun Kabir<sup>1,#</sup>, Md. Ershaduzzaman<sup>1</sup>, Md. Giasuddin<sup>1</sup>, Md. Rafiqul Islam<sup>1</sup>, K. H. M. Nazmul Hussain Nazir<sup>2</sup>, Mohammed Sirajul Islam<sup>1</sup>, Md. Rezaul Karim<sup>1</sup>, Md. Hafizur Rahman<sup>1</sup> and Md. Yousuf Ali<sup>1</sup>

• Received: Aug 29, 2017 • Revised: Sep 18, 2017 • Accepted: Sep 19, 2017 • Published Online: Sep 21, 2017



#### **AFFILIATIONS**

<sup>1</sup>Bangladesh Livestock Research Institute (BLRI), Savar, Dhaka, Bangladesh.

<sup>2</sup>Department of Microbiology and Hygiene, Bangladesh Agricultural University, Mymensingh-2202, Bangladesh.

#### ABSTRACT

**Objective:** The study was conducted to assess the prevalence of subclinical mastitis in dairy cows at Baghabari milk shed area of Sirajganj and Pabna districts in Bangladesh.

**Materials and methods:** A total of 300 milk samples were collected from crossbred dairy cows, and the milk samples were subjected for using California Mastitis Test (CMT). Besides, data related to farm management were taken from 60 farmers through direct interviews. The CMT was compared with White Slide Test (WST) and Surf Field Mastitis Test (SFMT). The samples showing strong positive reaction to mastitis by CMT were used for the isolation of associated bacteria using conventional bacteriological examinations and biochemical properties. The isolated bacteria were subjected for antibiogram studies by disc diffusion method.

**Results:** Out of 300 samples, 153 (51%) revealed positive reactions denoting that the cows were suffering from subclinical mastitis. Of the 153 samples, 39(13%) were trace-positive, 56(18.56%) were weak-positive, 33(11%) were distinct-positive, and the rest 25(8.3%) samples were strongly positive. In comparison economically among CMT, WST and SFMT, SFMT was found to be the cheapest and easiest. In this study, the cows were mostly infected with mixed infection with *Staphylococcus spp.*, *Streptococcus spp.*, *Escherichia coli, Salmonella spp.*, and some cows had single bacterial infection. It was observed that most of the microorganisms were sensitive to Gentamycin, Amoxicillin and Ceftriaxone, and were resistant to Penicillin, Ciprofloxacin and Colistin sulphate.

**Conclusion:** Overall prevalence of subclinical mastitis in the crossbred dairy cows at Sirajganj and Pabna district is 51%. It indicates that subclinical mastitis is a major threat for dairy industry in the studied areas. Early detection and appropriate preventive measures are suggestive to successful control of the disease.

### KEYWORDS

Bacteria; Cows; Sub-clinical mastitis; Prevalence

**How to cite:** Kabir MH, Ershaduzzaman M, Giasuddin M, Islam MR, Nazir KHMNH, Islam MS, Karim MR, Rahman MH, Ali MY. Prevalence and identification of subclinical mastitis in cows at BLRI Regional Station, Sirajganj, Bangladesh. Journal of Advanced Veterinary and Animal Research, 2017; 4(3):295-300.

## E-mail: <u>hkabirvet@yahoo.com</u>

Bangladesh Livestock Research Institute (BLRI), Savar, Dhaka, Bangladesh.

CORRESPONDENCE:

<sup>#</sup> Md. Humayun Kabir,

http://bdvets.org/javar/

#### INTRODUCTION

Mastitis is characterized by increased somatic cell count in the milk of affected animal. Subclinical mastitis shows no changes in the milk apparently. However, production of milk can be reduced up to 80% due to subclinical mastitis. If the disease persists for long time it adversely affects milk quality (Seegers et al., 2003). The prevalence of mastitis is mostly found in the farm condition with larger hard size as compared to those of lower herd sizes (Radostits et al., 2000). Mastitis may originate from microbial cause; the microbes may include virus, bacteria, mycoplasma and yeast (Egwu et al., 1994).

Although difficult as compared to clinical mastitis, early detection of subclinical mastitis in cow is crucial for saving dairy cattle and reduction of losses of farmers. Considering physical and chemical changes of milk and isolation of associated organisms, several methods have been developed to identify subclinical mastitis (Badiuzzaman et al., 2015). Among the tests, California Mastitis Test (CMT), White Side Test (WST) and Surf Field Mastitis Test (SFMT) are arguably the reliable screening tests for sub clinical mastitis (Greiner et al., 2000).

Staphylococci are the major etiological agents followed by Streptococci and Escherichia coli causing sub-clinical mastitis in cows in India (Singh and Baxi, 1982). In Bangladesh, clinical cases of mastitis caused by several bacteria like Staphylococci, Streptococci, Corynebacterium, E. coli and Bacillus sp. (Mahbub-E-Elahi et al., 1996). Subclinical mastitis is a serious problem for dairy industry as there are no gross changes found in udder or glandular tissues. Thus, it needs laboratory examination. Sub-clinically infected animal may act as a continuous source of infection to herd mates. If sub-clinical mastitis persists in dairy industry, it causes huge losses. The annual economic loss due to reduced milk production in Bangladesh estimated to be Taka 122.6 (US\$ 2.11) million (Kader et al., 2003). Considering the above situation, prevalence study of subclinical mastitis and identifying the causal agents are crucial. The objectives of the present study were to assess the prevalence of sub-clinical mastitis in cows in Sirajganj and Pabna districts of Bangladesh, to identify the causal agents, and to build awareness among the farmers.

#### MATERIALS AND METHODS

**Selection of study area, duration and study animal:** A total of 300 crossbred dairy cows were selected from 13 villages for this study and were sampled during the period of September 2015 to March 2016 from different dairy farms at Baghabari milk shed area of Sirajganj and Pabna

districts in Bangladesh. Number of cows per farm was between 3 and 50. In this study, only apparently healthy crossbred dairy cows were considered.

**Sample collection:** In this study, 300 samples were collected from Baghabari milk shed area of Sirajganj and Pabna districts. Before collection of milk, the teat and tips were washed with clean water, antisepsis was done with a swab soaked with 70% alcohol and then milk samples were collected aseptically from the udder during morning. All the milk samples were collected in vials which were labeled with identification number of cow.

**Physical examination of milk sample:** Immediately after collection, the milk samples were subjected to physical examination with naked eyes to detect any abnormalities in color, consistency and presence of any other clotted blood flakes and any other visible abnormalities.

**Detection of sub-clinical mastitis by CMT tester:** For detection of sub-clinical mastitis, CMT was performed as the instructions of manufacturer (CHEIL BIO Co. Ltd.). In brief, 2 mL milk and 2 mL CMT solution were mixed together in test paddle. Rotate the paddle to mix, and changes in color and gel formation was observed within 10 to 15 Sec.

**Transportation of milk sample:** The milk samples of strong positive results (+++) in CMT were transferred to Animal Health Laboratory, Bangladesh Livestock Research Institute (BLRI), Regional Station Baghabari, Sirajganj and also to Animal Health Division, BLRI, Head Quarter, Savar, Dhaka by cool box with ice.

Isolation of bacteria from milk: Before incubation, the sample was allowed in normal temperature. Then 100 µL of milk sample was taken with micro pipette and the sample was expelled on a test-tube containing 10 mL nutrient broth. These works were performed within a biohazard safety cabinet for aseptic measure. Then the milk sample containing broth was incubated at 37°C for 24 h. After incubation, one loop of incubated sample was streaked on EMB agar, Mac Conkey agar, Brilliant Green Agar and Mannitol Salt Agar respectively and again incubated at 37°C for 24-48 h. The bacterial isolates were identified by their cultural, morphological and biochemical characters (Rahman et al., 2014). The classification and specification of organism was based on the scheme presented in Bergey's Manual of Systemic Bacteriology (Halt et al., 1985).

Antibiogram studies: The isolated bacterial were subjected for antibiogram studies using commonly used antibiotics (Gentamycin 10  $\mu$ g/disc, Amoxicillin 20  $\mu$ g/disc, Ceftriaxone 30  $\mu$ g/disc, Penicillin G 10 units/disc, Ciprofloxacin 5  $\mu$ g/disc and Colistin sulphate 10  $\mu$ g/disc) following disc diffusion method, as described by Bauer et al. (1966).

#### **RESULTS AND DISCUSSION**

In this study, we found that farmers were practiced their animal deworming programs in different ways such as every 3, 6 and 9 months, 1 year and >1 year, and the percentage of it practicing farmers were 15, 50, 3.33, 26.67 and 5%, respectively. About 98 % farmers were not practicing of milking hand and udder washed out with antiseptic solution or water founded in this study. About 93 % farmers were not practicing of bathing of their dairy cows before milking and there were no certain hygienic place of milking. In this study, we found that 98% cow's sheds were tin shed and 76.67% floors were made up of brick. Hygienic status of housing and floor was not satisfactory for these dairy farms. Only 6.67% farmers used antiseptic in the cleaning program of their farms. They have no isolation shed for sick animals in their farms. Only 3.34% farmers adopted manure management system such as Biogas plant.

We found in this study that 54% dairy farmers claimed 30-60% reduced milk production due to mastitis in the affected cows. About 56.67% dairy farmers had previous record of culling of cows due to mastitis. Among the 300 cows, CMT was positive for 51% (trace- 13%, weak 11%, distinct 18.67%, strong 8.33%) animals. Among them, 100 were again screened with WST and SFMT showing negative 45 and 45%, mild 22 and 20%, moderate 27 and 29%, strong 6 and 6%, respectively. There are still factors

	Table 1	1: D	escription	of milk	quality
--	---------	------	------------	---------	---------

related to the sanitary management that may directly influence on SCC ( $\frac{1}{1200}$  et al., 2001).

Kader et al. (2002) reported similar report who described the prevalence as 44.61% sub-clinical mastitis in Bangladesh. On the other hand, higher prevalence (54%) of sub-clinical mastitis was recorded in India by Singh and Baxi (1982). As compared with early lactation (41.3%), prevalence of SCM was higher in late lactation period (58.5%), as reported by Radostits et al. (2000). Prevalence of sub-clinical mastitis was higher in high milk yielding animals (73%) as compared to low milk yielding animals (7.7%) (Khanal and Pandit, 2013). Previously, in Bangladesh, prevalence of sub-clinical mastitis in dairy cows was recorded as 72.07, 66.67, 64.86 and 61.26% by CMT, SCC, WST and SFMT, respectively, as reported by Badiuzzaman et al. (2015). The highest prevalence was found for CMT (67%) and WST (62%) in the animals aging 3.5-4.5 years (Khokon et al., 2017).

The result of this study was much higher than that of (Rahman et al., 2014) who reported only 20.13% in Friesian and more or less similar in case of Red Chittagong cows which was 13.24%. From India, (Devi et al., 1997) had reported 75.3% subclinical mastitis. This was also higher result than the present findings. Sargeant et al. (2001) studied the sensitivity of SCC and CMT for identifying intramammary infection during early lactation and found that CMT would better as compared to SCC. Barbosa et al. (2002) described that the SCC and CMT were dependent. Sudhan et al. (2005) reported that the percentage of agreement of CMT, SLST, bromothymol blue card test and WST with bacteriological examination

Reaction	Decision	Somatic cell	Polymorph				
		counts/mL	nuclearleukocyte (%)				
Mixture remain liquid state	-	0 ~ 200,000	0~25				
A slight slime formed , Trace reactions tend to	±	150,000~500,000	30~40				
disappear with continued movement of the fluid							
A distinct slime but with no tendency toward gel	+	400,000 ~1,500,000	40~60				
formation.							
The mixture thickens with gel formation, collected to	++	800,000~ over 5,000,000	60~70				
center of cups when rotate the test paddle.							
The mixture thickens with gel formation, icky adhere to	+++	Over 5,000,000	70~80				
base of cups. There is a central peak which remains							
projecting above the main mass after then motion of							
the paddle has been stopped. Viscosity is greatly							
increased so that there is a tendency for the mass to							
adhere to the bottom of the cup.							
Indicated by a contrasting deeper purple color.	Alkali milk	Over 5,000,000	Over 80				
Is's the result of inflammation or in drying-off of the							
mammary gland.							
Bromocresol purple is distinctly changes yellow color at	Acidic milk	Over 5,000,000	Over 80				
рН 5.2							

**Table 2:** Cultural colony characteristics and Gram's staining reaction of the organisms isolated from sub-clinical mastitis affected cows.

MCA	XLD	MSA	EMB	Gram stain	Arrangement	Tentative diagnosis
No growth	No growth	Medium-sized	No growth	+ve	Cluster	Staphylococcus spp.
		yellow colonies				
Rose pink lactose	No growth	No growth	Yellow green	-ve	Single paired or	Escherichia coli
fermented colony	-	-	metalic sheen		in short chain	
No growth	Black colony	No growth	No growth	-ve	Black spotted	Salmonella spp.

MCA=Mac Conkey Agar, XLD=Xylose Lysine Deoxycholate Agar, MSA=Mannitol Salt Agar, EMB=Eosin Methylene Blue agar



Figure 1. Prevalence of subclinical mastitis in dairy cows by CMT. Overall prevalence is 51%.

were 57.89, 62.07, 64.46 and 68.65%, respectively. This variation might be due difference in climatic condition, breed, management practices and treatment of cattle.

In this study, culture was done from the milk samples that demonstrated strong positive reaction in CMT. Mahbub-E-Elahi et al. (1996) isolated Staphylococcus, Streptococcus, E. coli and Bacillus, and Rahman et al. (1968) isolated and identified different strains of Staphylococci from mastitic and apparently healthy mammary glands of cows. These findings also corresponded with Shrestha and Bindari (2012) who reported highest prevalence of Staphylococcus followed by E. coli, Streptococci and Corynebaceterium. Chanda et al. (1998) reported that Staphylococcus was the principal organism of mastitis. Staphylococcus is the opportunistic bacterium which can survive the skin of the udder can infect via teat canal. In addition to Staphylococcus, another bacteria E. coli was identified in this study, which is an environmental opportunistic pathogen. Similar report was also reported by Mahbub-E-Elahi et al. (1996).

The occurrence of mastitis mainly depends on udder resistance, balance of pathogenic organisms and also

period of the exposure of the lactic gland to infection (<u>lanzekovic et al., 2009</u>).

The isolates of *Staphylococcus spp.* were found to ferment all the five basic sugars and produced only acid whereas the isolates of *E. coli* were found to ferment all the five basic sugars and produced both acid and gas. Culture sensitivity test was performed against Gentamycin, Amoxicillin, Ceftriaxone, Penicillin, Ciprofloxacin and Colistin sulphate. It was observed that most of the microorganisms were sensitive to Gentamycin, Amoxicillin, Ceftriaxone and showed resistant to Penicillin, Ciprofloxacin and Colistin sulphate.

#### CONCLUSION

The prevalence of subclinical mastitis indicate that it is the major threat for dairy industry Further study should be taken into consideration to identify the specific causal agents and develop a mastitis detection kit or mastitis control vaccine for better prevention of the disease. Ensure early detection and taken preventive measures immediately then it will be helpful to control the disease successfully.

#### ACKNOWLEDGEMENT

The authors thank the Bangladesh Livestock Research Institute (BLRI) for funding this research project. The authors also thank the farmers and workers for their participation and cooperation.

#### **CONFLICT OF INTEREST**

The authors have declared that there is no conflict of interest.

#### **AUTHORS' CONTRIBUTION**

HK, E, G and RI implemented the study design. MSI and YU participated in data collection and HK, also performed all the tests. HK and HR drafted, RK revised the manuscript. KHMNH critically checked the article and corrected the manuscript. All authors read and approved the final version of manuscript.

#### REFERENCES

- Badiuzzaman M, Samad MA, Siddiki SHMF, Islam MT, Saha S. Subclinical mastitis in lactating cows: comparison of four screening tests and effect of animal factors on its occurrence. Bangladesh Journal of Veterinary Medicine. 2015; 2:41-50. http://dx.doi.org/10.3329/bjvm.v13i2.26627
- Barbosa CB, Benadetti E, Ribeiro SC, Guimaraes EC, Ribeiro SC. The relationship between somatic cell count (SCC) and result of the California Mastitis Test (CMT) to diagnose bovine mastitis. Bioscience Journal. 2002; 18:93-102.
- Bauer A, Kirby W, Sherris JC, Turck M. Antibiotic susceptibility testing by a standardized single disk method. American Journal of Clinical Pathology. 1966; 45:493.
- Chanda A, Roy CR, Banerjee PK, Guha C. Studies of incidence of bovine mastitis, its diagnosis, etiology and in vitro sensitivity of the isolated pathogens. Indian Veterinary Journal. 1998; 66:277-282.
- Devi BK, Shukla PC, Bagherwal RK. Incidence of sub-clinical mastitis in cows. Indian Journal Dairy Science. 1997; 50:477-478.
- Egwu GO, Zaria LT, Onyeyili PA, Ambali AG, Adamu SS, Birdling M. Studies on the microbiological flora of caprine mastitis and antibiotic inhibitory concentrations in Nigeria. Small Ruminant Research. 1994; 14:233-239. https://doi.org/10.1016/0921-4488(94)90046-9
- 7. Greiner M, Pfeiffer D, Smith RD. Principles and practical application of the receiver-operating

characteristics analysis for diagnostic tests. Preventive Veterinary Medicine. 2000; 45:23-41. https://doi.org/10.1016/S0167-5877(00)00115-X

- Halt JG, Krieg NR, Sneath PH, Stely JJ, Williums ST. Bergey's Manual of Systemic Bacteriology. 1<sup>st</sup> Edn., London. 1985.
- Ítavo LCV, dos Santos GT, de Toledo VDA, Ítavo CCBF, Ribas NP. Milk quality and subclinical mastitis detection through somatic cells Counting. Acta Scientiarum. 2001; 4:1065-1068.
- Janzekovic M, Brus M, Mursec B, Vinis P, Stajnko D, Cus F. Mastitis detection based on electric conductivity of milk. Journal of Achievements in Materials and Manufacturing Engineering. 2009; 34(1):39-46.
- Kader MA, Samad MA, Saha S, Taleb MA. Prevalence and aetiology of sub-clinical mastitis with antibiotic sensitivity to isolated organisms among milch cows in Bangladesh. Indian Journal of Dairy Science. 2002; 55:218-223.
- Kader MA, Samad MA, Saha S. Influence of host level factors on prevalence and economics of subclinical mastitis in dairy cows in Bangladesh. Indian Journal of Dairy Science. 2003; 56:235-240.
- Khanal T, Pandit A. Assessment of sub-clinical mastitis and its associated risk factors in dairy livestock of Lamjung, Nepal. International Journal of Infection and Microbiology. 2013; 2(2):49-54. <u>https://doi.org/10.3126/ijim.v2i2.8322</u>
- Khokon MSI, Azizunnesa, Islam MM, Chowdhury KB, Rahman ML, Ali MZ. Effect of mastitis on postpartum conception of cross bred dairy cows in Chittagong district of Bangladesh. Journal of Advanced Veterinary and Animal Research. 2017; 4(2):155-160.

http://doi.org/10.5455/javar.2017.d203

- Mahbub-E-Elahi ATM, Rahman MA, Rahman MM, Prodhan MAM. Isolation and identification of bacteria from different quarters of mastitis affected dairy cows in Bangladesh. Bangladesh Veterinary Journal. 1996; 30:63-65.
- Radostits OR, Blood DC, Gay CC, Hinchcliff KW. Mastitis. In: Veterinary Mededicine, A textbook of the diseases of cattle, sheep, goats and horses. 8<sup>th</sup> Edn., Bailler Tindall, London. 2000; p. 603-700.
- Rahman MA, Chowdhury TIMFR, Chowdhury MUA. Distributing of different strains of *Staphylococcus* from mastitic and apperently normal bovine mammary gland. Pakistan Journal of Veterinary Science. 1968; 2:63-67.
- Rahman MM, Munsi MN, Ekram MF, Kabir MH, Rahman MT, Saha S. Prevalence of subclinical mastitis in cows at anwara, a coastal upazila of

Chittagong district in Bangladesh. Journal of Veterinary Advances. 2014; 6:594-598.

- Sargeant JM, Leslie KE, Shirley JE, Pulkrabek BJ, Lim GH. Sensitivity and specificity of somatic cell count and California mastitis test for identifying intramammary infection in early lactation. Journal of Dairy Science. 2001; 84:2018-2024. <u>https://doi.org/10.3168/jds.S0022-0302(01)74645-0</u>
- Seegers H, Fourichon C, Beaudeau F. Production effects related to mastitis and mastitis economics in dairy cattle herds. Veterinary Research. 2003; 34:475-491. <u>https://doi.org/10.1051/vetres:2003027</u>
- 21. Shrestha S, Bindari YR. Prevalence of sub-clinical mastitis among dairy cattle in Bhaktarpur District, Nepal. International Journal of Agriculture and Biosciences. 2012; 1(1):16-19.
- 22. Singh KB, Baxi KK. Studies on the etiology *in vitro* sensitivity and treatment of subclinical mastitis in milch animals. Indian Veterinary Journal. 1982; 59:191-198.
- Sudhan NA, Singh R, Singh M, Soodan JS. Studies on prevalence, etiology and diagnosis of subclinical mastitis among cross bred cows. Indian Journal of Animal Research. 2005; 39:127-130.

\*\*\*\*