

ORIGINAL ARTICLE

## The relationship between G1 (c.260 G>A) and G4 (c.721 G>A) polymorphisms in the GDF9 gene and the litter size of sheep: A meta-analysis study

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### ABSTRACT

**Objective:** The results of G1 and G4 polymorphisms as litter-size (LS) markers of ewes remain contradictory. The aim was to evaluate the impact of G1 (c.260 G>A) and G4 (c.721 G>A) polymorphisms on the LS of sheep by synthesizing data from multiple previous studies.

**Methods:** Data were extracted from 14 eligible articles. The genotypes of G1 and G4 polymorphisms were homozygous wild-type (WW), heterozygous (WM), and homozygous mutant-type (MM). The standardized mean difference (SMD) method using random effect models was employed to determine the effect size of G1 and G4 polymorphisms on LS under dominant, recessive, additive, and co-dominant genetic models. Heterogeneity was analyzed with the I<sup>2</sup> statistic index. Publication bias was depicted with funnel plots and tested by Egger's and Begg's tests.

**Results:** The study showed that the correlation between G1 polymorphism and LS in sheep was not significant ( $p > 0.05$ ) under all genetic models. The influence of G4 polymorphism on the LS of sheep was found significantly ( $p < 0.05$ ) under dominant [SMD = 0.28, I<sup>2</sup> = 0% (no heterogeneity)] and co-dominant [SMD = -0.14, I<sup>2</sup> = 36% (moderate heterogeneity)] genetic models. The WM genotype of G4 polymorphism increased LS, while the MM genotype reduced LS in sheep. Publication bias among G1 and G4 polymorphism studies was absent in all genetic models.

**Conclusion:** Thus, the study revealed that G4 polymorphism could be a potential genetic marker for LS in ewes. On the contrary, G1 polymorphism has no association with the LS of ewes.

### ARTICLE HISTORY

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### KEYWORDS

Fecundity Gene; Litter Size;  
Ovis aries; Single Nucleotide  
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### Introduction

Domesticated sheep are small ruminants with essential values in society's lives [1], such as providing food and wool, economic sources, cultural and religious festivities, and other roles [2,3]. Reproductive traits play a critical role in the sheep industry [4]. They have an impact on sustainability and profitability [5,6]. Litter size (LS), the number of offspring born to a female animal at a single birth, is one of the crucial reproductive traits of livestock production, especially in the sheep industry [7]. It affects the profitability and efficiency of sheep production [8]. The increment in LS from 1.0 to 2.2 lambs per ewe is linear with the increase in gross margin revenue in the sheep industry [9]. Genetic factors are fundamental for LS, besides nutrition

and environmental conditions. Several fecundity genes have been identified as valuable genetic markers because of their effect on the LS of ewes [10]. Exploring specific genes as LS markers is necessary to enhance sheep's productivity.

Growth differentiation factor 9 (*GDF9*) is renowned for crucially regulating ovarian follicle development in mammals, including the growth of oocytes and granulosa cells [11] and theca cell proliferation [12]. Oocytes primarily secrete *GDF9* [13]. *GDF9* has crucial functions in ovulation and fertilization [14]. In sheep, the *GDF9* gene was found on chromosome 5 [15], which comprises two exons and one intron. The *GDF9* gene has been identified as the putative marker for the prolificacy of sheep and goats [16–19].

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*G1* (c.260 G>A) and *G4* (c.721 G>A) polymorphisms in the *GDF9* gene are the most studied loci in the *GDF9* gene used to investigate the fertility traits of sheep based on the prolificacy parameter. *G1* causes the alteration of arginine for histidine (*R87H*) at coding residue 87 in exon 1 and *G4* changes glutamic acid to lysine (*E241K*) at coding residue 241 [20,21]. However, the results of *G1* and *G4* polymorphisms on the LS of ewes remain contradictory. Several studies reported that *G1* and *G4* polymorphisms have affected the LS of sheep [22–29]. Meanwhile, there are other investigations that discovered no association between *G1* and *G4* polymorphisms and LS of sheep [30–34]. Therefore, advanced statistical analysis is required to elucidate the impact of *G1* and *G4* polymorphisms on the LS of ewes based on the findings of previous studies.

Meta-analysis is a systematic study designed to statistically synthesize the data of multiple previous studies to derive a quantified conclusion [35]. It enables researchers to integrate the data from insufficiently powered experiments to generate conclusive findings [36]. Because of a data-pooled study, the result of a meta-analysis study may be more precise and robust than the individual study [37]. Numerous meta-analysis studies have been employed to assess the influence of the fecundity genes *BMP15* and *BMPR1B* polymorphisms on the LS of sheep and goats [38,39] and the relationship between c.1189 G>A variation in the *GDF9* gene and the LS of dose [40]. To the best of our investigation, no meta-analysis examining the correlation between the *G1* and *G4* polymorphisms in the *GDF9* gene and the LS of sheep has been found. Thus, the aim was to evaluate the link between *G1* and *G4* polymorphisms in the *GDF9* gene and the LS of sheep by synthesizing data from numerous published articles.

## Materials and Methods

### Literature search strategy

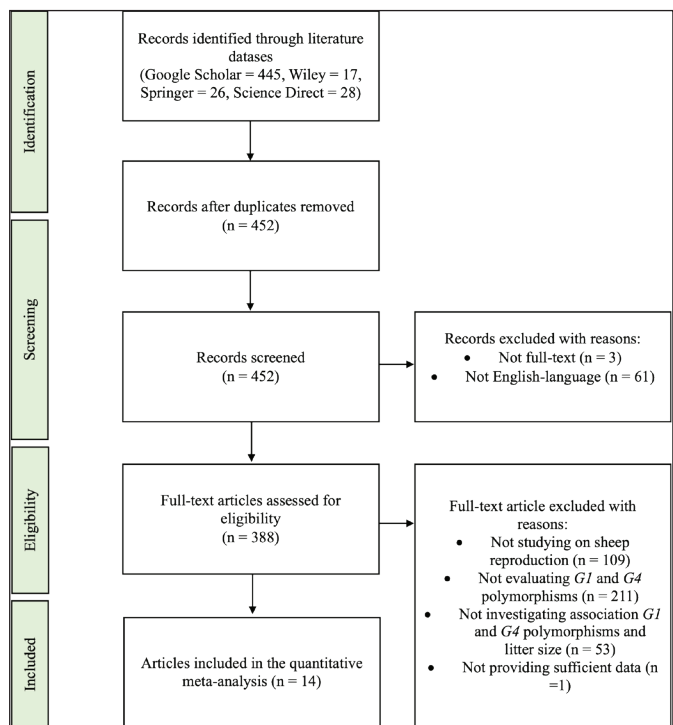
The meta-analysis study used preferred reporting items for systematic reviews and meta-analyses (PRISMA) guidelines to qualify the eligible studies [41]. Multiple academic databases, including Google Scholar, Science Direct, Wiley, and Springer, were independently employed to retrieve the related studies by two scientists (AB and SH) with several keywords, alone or in combination, as follows: “*GDF9*,” “Polymorphism,” “SNP,” “Prolificacy,” “Fertility,” “LS,” and “Sheep.” Moreover, we verified that no articles were missed by scrutinizing the reference list of extracted articles. All differences concerning the eligibility of the studies were settled through discussions. Finally, if no agreement regarding the inclusion of eligible studies was obtained, a discussion with a third scientist (RW) was used to resolve all remaining differences.

### Inclusion and exclusion criteria

The eligible studies were included if they were: (1) studying *G1* (c.260 G>A) and *G4* (c.721 G>A) polymorphisms in sheep; (2) each genotype is provided with the number of samples; (3) each genotype displays the least-squares mean (LSM); (4) evaluating the correlation *G1* and *G4* polymorphisms and LS; and (5) reporting a standard error (SE) or standard deviation (SD) or confidence interval (CI). The following criteria apply to excluded studies: (1) non-English-language articles; (2) no full text is available; (3) relevant data are insufficient; (4) duplicated studies; and (4) review studies.

### Data extraction

Selected studies were extracted and input as follows: the first author’s name, publication’s year, location of the study, sheep breed, the number of samples, LSM, SD, and significant level. An online extractor, WebPlotDigitizer (<https://apps.automeris.io/wpd/>), was applied to extract the LSM and SE from the graphed data [42]. To obtain SD from sample sizes and SE of LSM from each genotype, the following formula was employed:  $SD = SE \cdot \sqrt{N}$ , where  $N$  is the number of samples from each genotype. Meanwhile, SD from the number of samples and 95% CI from each genotype were



**Figure 1.** The flow chart based on PRISMA guidelines illustrates the comprehensive process for screening included and excluded studies.

computed with the following formula:  $SD = x$  (upper limit—lower limit)/3.92 [43].

### Statistical analysis

The study used Review Manager v5.4 software and the “meta” package of R v4.2.2 software [44]. Data were synthesized with the standardized mean difference (SMD) method [41] using random-effects models under dominant ( $WW + WM$  vs.  $MM$ ), recessive ( $WW$  vs.  $WM + MM$ ), additive ( $WW$  vs.  $MM$ ), and co-dominant ( $WW + MM$  vs.  $WM$ ) genetic models to determine the effect size of  $G1$  and  $G4$  polymorphisms on the LS of ewes. The  $p$ -values  $<0.05$  indicated the effect was considered to be significant.

To test heterogeneity among studies, the  $I^2$  statistic index was applied. The  $I^2$  values  $<25\%$ ,  $I^2$  values between  $25\%$  and  $50\%$ , and  $I^2$  values  $>50\%$  indicate low, moderate, and high heterogeneity, respectively [45]. Moreover, we performed a sensitivity test to investigate the firmness of all results by erasing a single study of each genetic model at a time.

Publication bias among the studies was depicted using funnel plots and tested using Egger’s and Begg’s tests. The

presence of an asymmetric funnel plot and the  $p$ -value  $\leq 0.05$  of Egger’s and Begg’s demonstrate the high risk of publication bias [46].

## Results

### Characteristics of included eligible studies

The present study applied a PRISMA flow diagram to display the comprehensive process of selecting qualified research articles for a meta-analysis, as illustrated in Figure 1. A comprehensive search of multiple literature databases identified a total of 516 articles. After eliminating 64 duplicate papers, the remaining 452 publications were screened to eliminate those that did not provide full-text articles and no English-language articles, resulting in 388 articles for the eligibility assessment.

After applying the exclusion criteria, 373 articles were removed. One study was rejected because it did not provide SE, SD, or CI. Finally, 14 eligible articles were selected for the meta-analysis study. Table 1 displays the characteristics of eligible studies for our meta-analysis study.

**Table 1.** The characteristics of included studies in this meta-analysis study.

Study	Year	Country	Breed	Total sample	Genotypes <sup>a</sup>			LSM $\pm$ SD <sup>b</sup>			Sig.
					WW	WM	MM	WW	WM	MM	
<i>G1</i> (c.260 G>A) polymorphism											
Abdelgadir et al. [22]	2021	Sudan	Sudanese Desert	88	52	30	6	1.248 $\pm$ 0.238	1.386 $\pm$ 0.252	1.040 $\pm$ 0.230	yes
Aboelhassan et al. [23]	2021	Egypt	Egyptian	95	92	3	NE	2.170 $\pm$ 4.220	0.048 $\pm$ 0.064	NE	yes
Hossain et al. [50]	2020	Bangladesh	Bangladesh	126	65	57	4	1.590 $\pm$ 0.726	1.830 $\pm$ 0.755	2.000 $\pm$ 0.820	yes
Rezaei et al. [26]	2020	Iran	Iran-Black	114	63	45	6	1.650 $\pm$ 0.714	1.890 $\pm$ 0.604	1.880 $\pm$ 0.392	yes
Pineda et al. [33]	2018	Colombia	Colombia Hair	150	123	27	NE	1.240 $\pm$ 0.300	1.330 $\pm$ 0.360	NE	no
Talebi et al. [34]	2018	Iran	Mehraban	115	82	31	2	1.160 $\pm$ 0.873	1.170 $\pm$ 0.856	1.000 $\pm$ 0.252	no
Eghbalsaied et al. [31]	2017	Iran	Afshari	145	118	20	7	1.082 $\pm$ 0.554	1.326 $\pm$ 0.936	1.005 $\pm$ 0.027	no
Eghbalsaied et al. [31]	2017	Iran	Ghezel	126	93	30	3	1.611 $\pm$ 0.779	1.498 $\pm$ 0.767	1.999 $\pm$ 0.019	no
Eghbalsaied et al. [31]	2017	Iran	Lori-Bakhtiyari	171	153	18	NE	1.578 $\pm$ 0.819	1.772 $\pm$ 0.609	NE	no
Eghbalsaied et al. [31]	2017	Iran	Shal	54	44	5	5	1.549 $\pm$ 0.793	1.331 $\pm$ 0.755	1.668 $\pm$ 0.732	no
Gorlov et al. [24]	2018	Russia	Salsk	500	440	60	NE	1.130 $\pm$ 1.888	1.800 $\pm$ 0.929	NE	yes
Gorlov et al. [24]	2018	Russia	Volgograd	500	420	80	NE	1.220 $\pm$ 2.254	1.180 $\pm$ 1.521	NE	yes
Jawasreh et al. [28]	2017	Jordan	Romanov	70	41	29	NE	2.732 $\pm$ 1.088	1.940 $\pm$ 0.969	NE	yes
Abdoli et al. [30]	2013	Iran	Mehraban	85	24	53	8	1.245 $\pm$ 0.315	1.103 $\pm$ 0.339	1.047 $\pm$ 0.205	no
Liandris et al. [32]	2012	Greece	Chios	239	156	62	21	1.590 $\pm$ 2.403	1.450 $\pm$ 1.615	2.250 $\pm$ 1.076	yes
Liandris et al. [32]	2012	Greece	Karagouniki	259	250	9	NE	1.320 $\pm$ 1.766	1.070 $\pm$ 0.507	NE	no
Moradband et al. [25]	2011	Iran	Baluchi	134	96	27	11	1.238 $\pm$ 0.314	1.386 $\pm$ 0.265	1.032 $\pm$ 0.312	yes
Roy et al. [27]	2011	India	Bonpala	97	86	10	1	1.670 $\pm$ 0.620	1.900 $\pm$ 0.570	1.000 $\pm$ 0.000	yes
Javanmard et al. [29]	2011	Iran	Fat-tailed	96	58	38	NE	1.160 $\pm$ 0.381	1.780 $\pm$ 0.432	NE	yes

(Continued)

Study	Year	Country	Breed	Total sample	Genotypes <sup>a</sup>			LSM ± SD <sup>b</sup>			Sig.
					WW	WM	MM	WW	WM	MM	
<i>G4</i> (c.721 G>A) polymorphism											
Aboelhassan et al. [23]	2021	Egypt	Egyptian	95	21	74	NE	0.300 ± 0.825	1.950 ± 4.731	NE	yes
Eghbalsaied et al. [31]	2017	Iran	Afshari	145	136	9	NE	1.240 ± 0.854	1.004 ± 0.031	NE	no
Eghbalsaied et al. [31]	2017	Iran	Ghezel	126	79	38	9	1.559 ± 0.858	1.665 ± 0.892	1.665 ± 0.985	no
Eghbalsaied et al. [31]	2017	Iran	Lori-Bakhtiyari	171	105	49	17	1.560 ± 0.768	1.520 ± 0.630	1.330 ± 0.660	no
Eghbalsaied et al. [31]	2017	Iran	Shal	54	28	24	2	1.753 ± 0.668	1.389 ± 0.569	1.500 ± 0.693	no
Gorlov et al. [24]	2018	Russia	Salsk	500	NE	60	440	NE	1.800 ± 0.930	1.130 ± 1.888	yes
Gorlov et al. [24]	2018	Russia	Volgograd	500	NE	80	420	NE	1.880 ± 1.521	1.220 ± 2.254	yes
Liandris et al. [32]	2012	Greece	Chios	239	154	79	6	1.830 ± 2.136	2.170 ± 1.518	1.300 ± 0.705	no
Liandris et al. [32]	2012	Greece	Karagouniki	259	197	49	13	1.130 ± 1.779	1.200 ± 0.975	1.260 ± 0.629	no
Talebi et al. [34]	2018	Iran	Mehraban	115	78	33	4	1.180 ± 0.601	1.160 ± 0.582	1.000 ± 0.236	no
Roy et al. [27]	2011	India	Bonpala	97	86	10	1	1.670 ± 0.620	1.900 ± 0.570	1.000 ± 0.000	no

<sup>a</sup>WW, homozygous wild-type genotype; WM, heterozygous genotype; MM, homozygous mutant-type genotype, <sup>b</sup>LSM, least-squares mean; SD, standard deviation; NE, no existence; Sig., significance.

**Table 2.** Summary of results of the meta-analysis on the relationship between *G1* (c. 260 G>A) and *G4* (c.721 G>A) polymorphisms in the *GDF9* gene and LS of sheep.

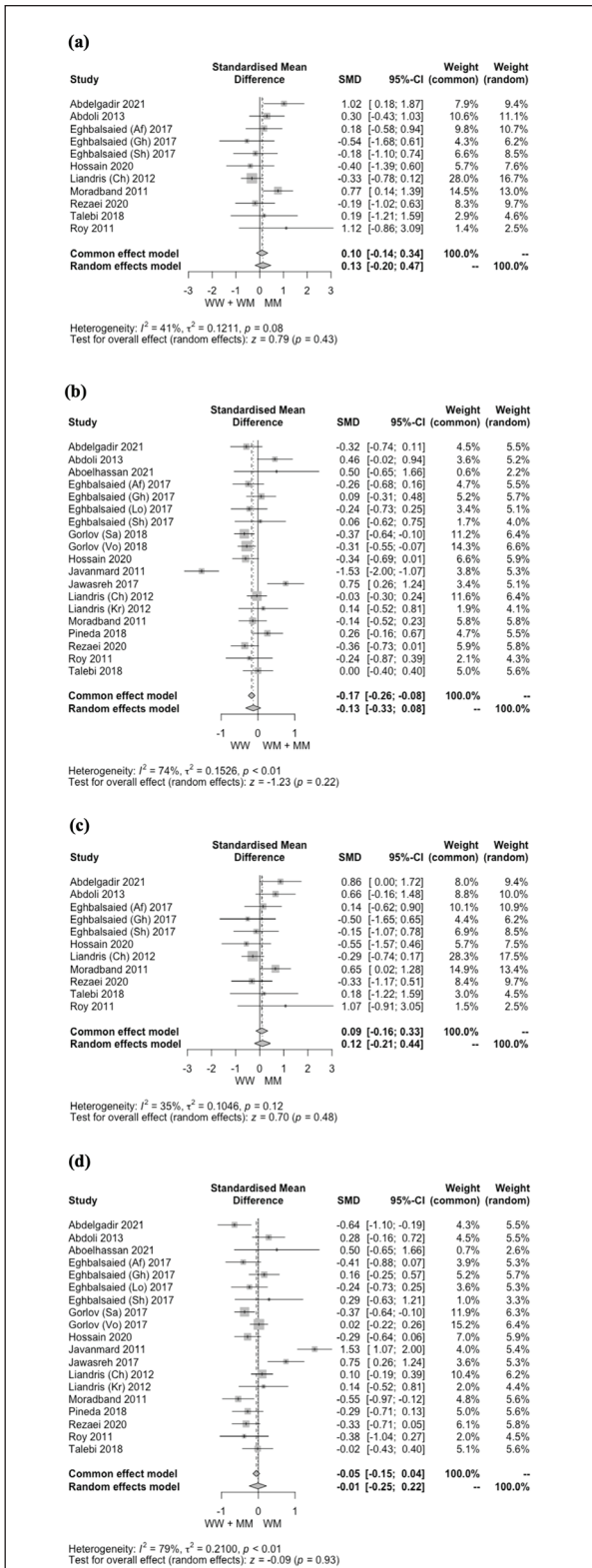
SNP <sup>a</sup>	Genetic model <sup>b</sup>	Number of cohort	Test of relationship <sup>c</sup>			Test of heterogeneity <sup>d</sup>		
			SMD	95% CI	p-value	Model	p-value	I <sup>2</sup>
<i>G1</i> (c.260 G>A)	Dominant (WW + WM vs. MM)	11	0.13	-0.20; 0.47	ns	R	0.08	41%
	Recessive (WW vs. WM + MM)	19	-0.13	-0.33; 0.08	ns	R	<0.0001	74%
	Additive (WW vs. MM)	11	0.12	-0.21; 0.44	ns	R	0.12	35%
	Co-dominant (WW + MM vs. WM)	19	-0.01	-0.25; 0.22	ns	R	<0.0001	79%
<i>G4</i> (c.721 G>A)	Dominant (WW + WM vs. MM)	9	0.28	0.13; 0.43	<0.01	R	0.87	0%
	Recessive (WW vs. WM + MM)	9	-0.03	-0.15; 0.10	ns	R	0.26	20%
	Additive (WW vs. MM)	7	0.14	-0.14; 0.43	ns	R	0.86	0%
	Co-dominant (WW + MM vs. WM)	11	-0.14	-0.27; -0.01	0.04	R	0.11	36%

<sup>a</sup>SNP, single nucleotide polymorphism, <sup>b</sup>WW, homozygous wild-type genotype; WM, heterozygous genotype; MM, homozygous mutant-type genotype, <sup>c</sup>SMD, standardized mean difference; CI, confidence interval. <sup>d</sup>R, random-effect models.

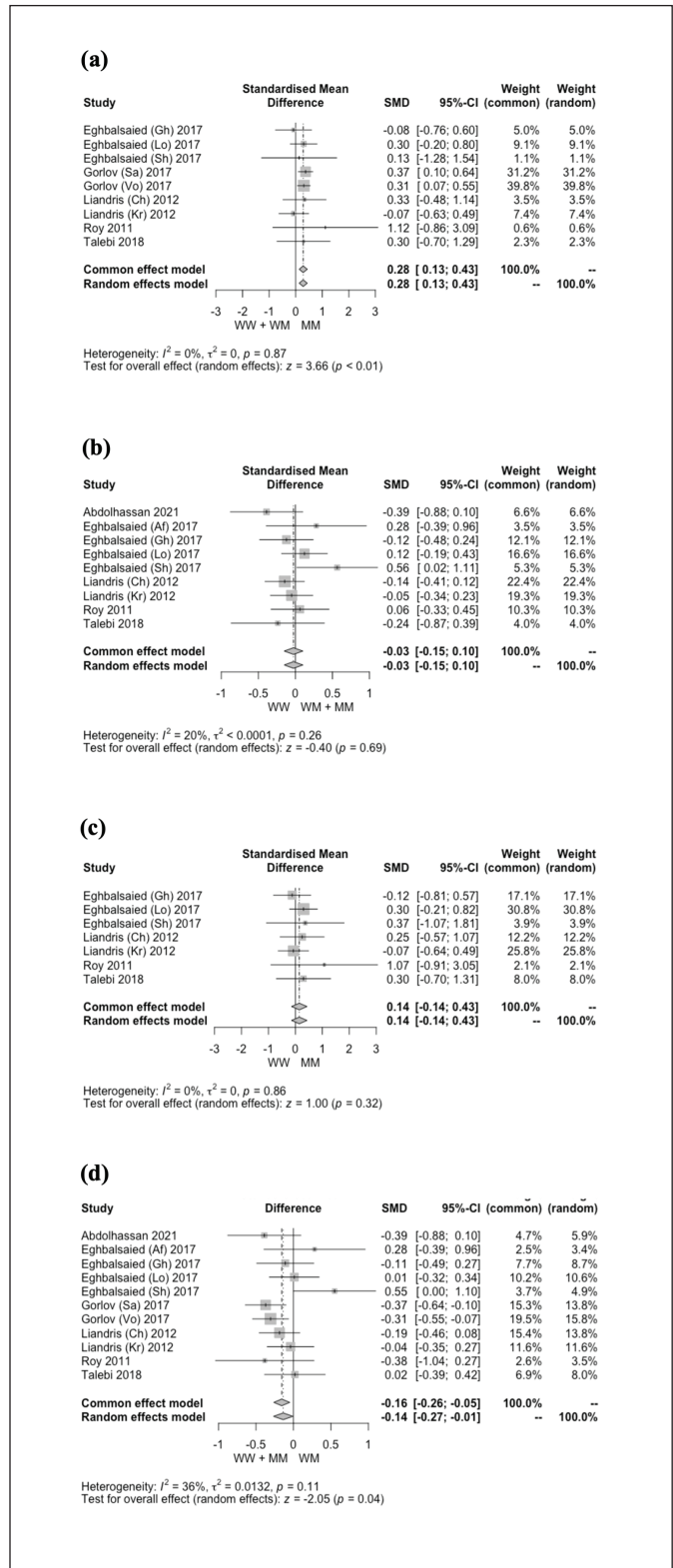
### Correlation between the *G1* and *G4* polymorphisms and LS of sheep

A comprehensive summary of the meta-analysis outcomes about the relationship between *G1* and *G4* polymorphisms in the *GDF9* gene and the LS of sheep is shown in Table 2. Figures 2 and 3 display the forest plots, illustrating meta-analysis findings on the effect of *G1* and *G4* polymorphisms on the LS of sheep. The result revealed

that *G1* polymorphism had no significant effect ( $p > 0.05$ ) on the LS of sheep under all genetic models. On the contrary, the LS of sheep was significantly affected ( $p < 0.05$ ) by *G4* polymorphism under dominant (SMD = 0.28) and co-dominant (SMD = -0.14) genetic models. In addition, the effect of *G4* polymorphism on LS was not observed ( $p > 0.05$ ) under recessive (SMD = -0.03) and additive (SMD = 0.14) genetic models.



**Figure 2.** Forest plot showing the association between *G1* (c.260 G>A) polymorphism in the *GDF9* gene and LS of sheep under dominant (a), recessive (b), additive (c), and co-dominant (d) genetic models.



**Figure 3.** Forest plot displaying the relationship between *G4* (c.721 G>A) polymorphism in the *GDF9* gene and LS of sheep under dominant (a), recessive (b), additive (c), and co-dominant (d) genetic models.

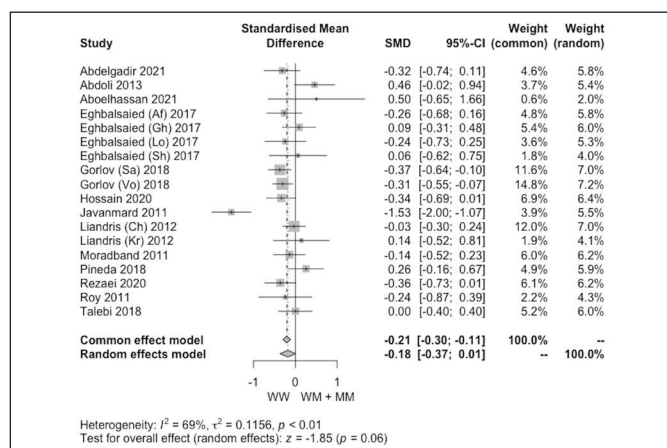
### Heterogeneity, sensitivity, and publication bias analysis

Moderate ( $25\% < I^2 \leq 50\%$ ) to high ( $I^2 > 50\%$ ) heterogeneity was found among *G1* polymorphism studies. The  $I^2$ -value under dominant, recessive, additive, and co-dominant genetics models among *G4* polymorphism studies was 0% (no heterogeneity), 20.1% (low heterogeneity), 0% (no heterogeneity), and 36% (moderate heterogeneity), respectively. Systematically erasing one study at a time and applying it to all models was the sensitivity test used to clarify the robustness of the pooled effect size. A significant change was found under the dominant genetic model in the *G1* study after removing Jawasreh et al.'s [28] reported *G1* polymorphism in Egyptian sheep, as shown in Figure 4. No significant change under all genetic models was found in the *G4* polymorphism study.

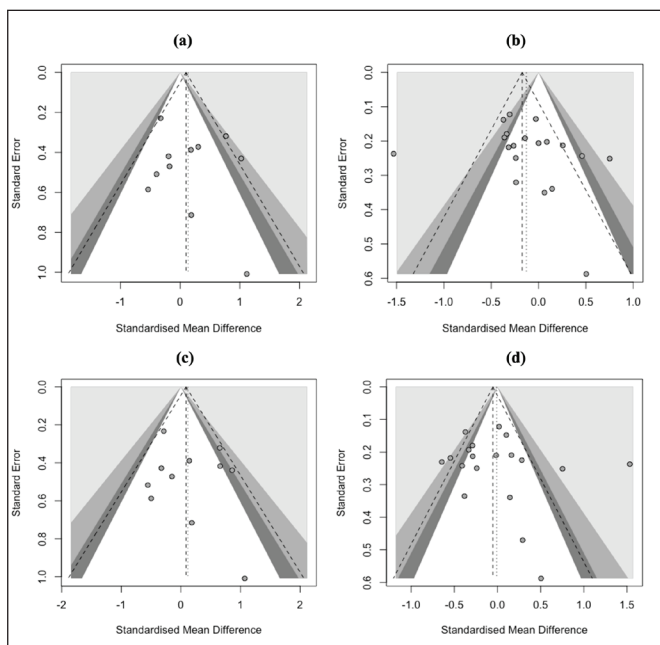
Figures 5 and 6 display the results of the publication bias test, as depicted by the funnel plots. No asymmetrical plots in all genetic models indicated publication bias was absent in the *G1* and *G4* studies. Egger's and Begg's tests were employed for all genetic models to confirm the funnel plot results. A significant publication bias in all models was also not found ( $p > 0.05$ ) in both studies, as shown in Table 3.

### Discussion

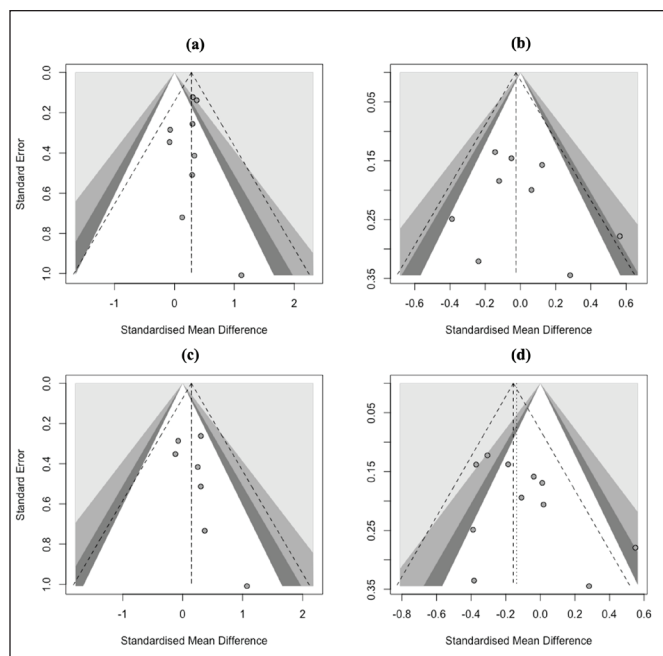
The *GDF9* gene has been identified as a highly prolific gene in sheep, along with *BMP15* and *BMP1B* [10]. *GDF9* gene single nucleotide polymorphisms (SNPs) affect the fertility traits of sheep [47]. The *G1* polymorphism (*c.260 G>A*) and *G4* (*c.721 G>A*) polymorphisms in the *GDF9* gene have been intensively studied to characterize the LS of sheep. However, the reported findings are still conflicting. Abdelgadir et al. [22] demonstrated that the *G1*



**Figure 4.** Forest plot showcasing sensitivity analysis of the effect of *G1* (*c.260 G>A*) polymorphism on LS of sheep under the recessive genetic model after removing Jawasreh et al. [28] studies (compared with Fig. 2b).



**Figure 5.** Funnel plots exhibiting analysis of publication bias among *G1* (*c.260 G>A*) polymorphism studies under the dominant model (a), recessive model (b), additive model (c), and co-dominant model (d).



**Figure 6.** Funnel plots depicting analysis of publication bias among *G4* (*c.721 G>A*) polymorphism studies under the dominant model (a), recessive model (b), additive model (c), and co-dominant model (d).

polymorphism effect significantly affected the LS of the Sudanese desert sheep. Moreover, Aboelhasan et al. [23]

**Table 3.** Summary of the risk of publication bias detected by Egger's and Begg's tests.

SNP <sup>a</sup>	Genetic model <sup>b</sup>	I <sup>2</sup>	p-value <sup>c</sup>	
			Egger's test	Begg's test
G1 (c.260 G>A)	Dominant (WW + WM vs. MM)	41%	0.56	0.69
	Recessive (WW vs. WM + MM)	74%	0.33	0.06
	Additive (WW vs. MM)	35%	0.58	0.82
	Co-dominant (WW + MM vs. WM)	79%	0.48	0.31
G4 (c.721 G>A)	Dominant (WW + WM vs. MM)	0%	0.62	0.83
	Recessive (WW vs. WM + MM)	20%	0.52	0.53
	Additive (WW vs. MM)	0%	0.22	0.18
	Co-dominant (WW + MM vs. WM)	36%	0.12	0.12

<sup>a</sup>SNP, single nucleotide polymorphism, <sup>b</sup>WW, homozygous wild-type genotype; WM, heterozygous genotype; MM, homozygous mutant-type genotype. <sup>c</sup>p-value >0.05, no publication bias; p-value <0.05, publication bias high.

and Gorlov et al. [24] demonstrated that the correlation between G1 and G4 mutations and LS in ewes was significant. However, Abdoli et al. [30], Eghbalsaied et al. [31], and Talebi et al. [34] revealed that the G1 and G4 polymorphisms had no association with the LS of sheep.

To settle the contradictory study results, meta-analysis has the ability to improve the precision and accuracy of scientific results by integrating and synthesizing pooled data from multiple studies. It provides robust and reliable conclusions that have positive impacts on many scientific studies [48]. Numerous studies have employed meta-analysis to evaluate the impact of fecundity gene polymorphisms on the LS of sheep and goats [39–41]. However, to the best of our knowledge, this study was the first comprehensive meta-analysis to investigate the association between G1 and G4 polymorphisms and the LS of sheep. We synthesized data from 14 articles under dominant, co-dominant, recessive, and additive genetic models.

This study demonstrated that the G1 polymorphism had no significant effect on the LS of sheep. However, the G4 mutation affected the LS, whereby the homozygous mutant-type (MM) genotype reduced the LS and the heterozygous (WM) genotype increased the LS of ewes. It is assumed that the heterozygous genotype of the G4 polymorphism in the GDF9 gene results in high ovulation rates, leading to higher LS. Hanrahan et al. [21] reported that the heterozygous genotype of the GDF9 gene boosts the ovulation rate. This finding was similar to the meta-analysis study conducted

by Mahmoudi et al. [40] reported that c. 1118 G>A polymorphism of the GDF9 gene had significantly affected the LS of goats, whereby the heterozygous genotype had a positive effect on LS.

Furthermore, no publication bias and moderate heterogeneity under the co-dominant genetic model assure an accurate estimate of the effect size in this study. No publication bias and low heterogeneity confirmed reliable effect sizes in the meta-analysis study [49].

However, it is assumably necessary to re-confirm our findings in future research because the present study has limitations. The several limitations of this study are as follows: a) the small number of included studies; b) the sample size was limited; and c) there was no sub-analysis based on the region of the sheep breed. The future study will focus on adding larger sample sizes to address the limitations of this study.

## Conclusion

Thus, the study revealed that G4 polymorphism could be a potential genetic marker for LS in ewes. On the contrary, G1 polymorphism has no association with the LS of ewes. However, the limitations of this study should be considered for further studies.

## List of abbreviations

BMP15, bone morphogenetic protein 15; BMPR1B, bone morphogenetic protein receptor type-1B; CI, confidence interval; G1, c.260 G>A mutation point in GDF9 gene; G4, c.721 G>A mutation point in GDF9 gene; GDF9, growth differentiation factor 9; I<sup>2</sup>, I statistic index LS, litter size; LSM, least mean square; MM, homozygous mutant-type genotype; PRISMA, Preferred reporting items for systematic reviews and meta-analyses; R, random-effect models; SD, standard deviation; SE, standard error; SMD, standardized mean difference; SNP, single nucleotide polymorphism; WM, heterozygous genotype; WW, homozygous wild-type genotype.

## Conflict of interests

All authors have no conflict of interest to declare.

## Authors' contributions

AB: first author, designed the study, screened literature databases for included studies, and revised the manuscript. SH: screened literature databases for included studies, analyzed data, and revised the manuscript. RW and EMNS: third-party discussion (screened included studies) and reviewed the manuscript. HPW and SG: extracted data, input data, and scrutinized missed data.

## References

- [1] Ibrahim A, Baliarti E, Budisatria IGS, Artama WT, Widayanti R, Maharani D, et al. Genetic diversity and relationship among Indonesian local sheep breeds on Java Island based on mitochondrial cytochrome b gene sequences. *J Genet Eng Biotechnol* 2023; 21(1):34; <https://doi.org/10.1186/s43141-023-00491-z>
- [2] Ibrahim A, Budisatria IGS, Widayanti R, Artama WT. Consumer's preferences for sheep attributes for Eid al-Adha celebration in Yogyakarta, Indonesia. *IOP Conf Ser: Earth Environ Sci* 2019; 387:012001; <https://doi.org/10.1088/1755-1315/387/1/012001>
- [3] Ibrahim A, Budisatria IGS, Widayanti R, Artama WT. The impact of religious festival on roadside livestock traders in urban and peri-urban areas of Yogyakarta, Indonesia. *Vet World* 2019; 12(9):1408–15; <https://doi.org/10.14202/vetworld.2019.1408-1415>
- [4] Ajafar HM, Kadhim HA, AL-Thuwaini MT. The reproductive traits of sheep and their influencing factors. *RAS* 2022; 10:82–9; [https://doi.org/10.7831/ras.10.0\\_82](https://doi.org/10.7831/ras.10.0_82)
- [5] Davis TC, White RR. Breeding animals to feed people: the many roles of animal reproduction in ensuring global food security. *Theriogenology* 2020; 150:27–33; <https://doi.org/10.1016/j.theriogenology.2020.01.041>
- [6] Budisatria IGS, Guntoro B, Sulfiar AET, Ibrahim A, Atmoko BA. Reproductive management and performances of Bali cow kept by smallholder farmers level with different production systems in South Konawe Regency, Indonesia. *IOP Conf Ser: Earth Environ Sci* 2021; 782:022079; <https://doi.org/10.1088/1755-1315/782/2/022079>
- [7] Tao L, He XY, Wang FY, Pan LX, Wang XY, Gan SQ, et al. Identification of genes associated with litter size combining genomic approaches in Luzhong mutton sheep. *Anim Genet* 2021; 52(4):545–9; <https://doi.org/10.1111/age.13078>
- [8] Zhang Z, Sui Z, Zhang J, Li Q, Zhang Y, Wang C, et al. Identification of signatures of selection for litter size and pubertal initiation in two sheep populations. *Animals* 2022; 12(19):2520; <https://doi.org/10.3390/ani12192520>
- [9] Farrell L, Creighton P, Bohan A, McGovern F, McHugh N. Bio-economic modelling of sheep meat production systems with varying flock litter size using field data. *Animal* 2022; 16(10):100640; <https://doi.org/10.1016/j.animal.2022.100640>
- [10] Liu Q, Pan Z, Wang X, Hu W, Di R, Yao Y, et al. Progress on major genes for high fecundity in ewes. *Front Agr Sci Eng* 2014; 1(4):282–90; <https://doi.org/10.15302/J-FASE-2014042>
- [11] Cook-Andersen H, Curnow KJ, Su HI, Chang RJ, Shimasaki S. Growth and differentiation factor 9 promotes oocyte growth at the primary but not the early secondary stage in three-dimensional follicle culture. *J Assist Reprod Genet* 2016; 33(8):1067–77; <https://doi.org/10.1007/s10815-016-0719-z>
- [12] Spicer LJ, Aad PY, Allen DT, Mazerbourg S, Payne AH, Hsueh AJ. Growth differentiation factor 9 (GDF9) stimulates proliferation and inhibits steroidogenesis by bovine theca cells: influence of follicle size on responses to GDF9. *Biol Reprod* 2008; 78(2):243–53; <https://doi.org/10.1095/biolreprod.107.063446>
- [13] Alam MH, Miyano T. Interaction between growing oocytes and granulosa cells *in vitro*. *Reprod Med Biol* 2020; 19(1):13–23; <https://doi.org/10.1002/rmb.2.12292>
- [14] Sanfins A, Rodrigues P, Albertini DF. GDF-9 and BMP-15 direct the follicle symphony. *J Assist Reprod Genet* 2018; 35(10):1741–50; <https://doi.org/10.1007/s10815-018-1268-4>
- [15] Våge DI, Husdal M, Kent MP, Klemetsdal G, Boman IA. A missense mutation in growth differentiation factor 9 (GDF9) is strongly associated with litter size in sheep. *BMC Genet* 2013; 14(1):1–8; <https://doi.org/10.1186/1471-2156-14-1>
- [16] Feng T, Geng CX, Lang XZ, Chu MX, Cao GL, Di R, et al. Polymorphisms of caprine GDF9 gene and their association with litter size in Jining Grey goats. *Mol Biol Rep* 2011; 38(8):5189–97; <https://doi.org/10.1007/s11033-010-0669-y>
- [17] Souza CJ, McNeilly AS, Benavides MV, Melo EO, Moraes JC. Mutation in the protease cleavage site of GDF9 increases ovulation rate and litter size in heterozygous ewes and causes infertility in homozygous ewes. *Anim Genet* 2014; 45(5):732–9; <https://doi.org/10.1111/age.12190>
- [18] Tong B, Wang J, Cheng Z, Liu J, Wu Y, Li Y, et al. Novel variants in GDF9 gene affect promoter activity and litter size in Mongolia sheep. *Genes* 2020; 11(4):375; <https://doi.org/10.3390/genes11040375>
- [19] Wang F, Chu M, Pan L, Wang X, He X, Zhang R, et al. Polymorphism detection of GDF9 gene and its association with litter size in Luzhong Mutton sheep (*Ovis aries*). *Animals* 2021; 11(2):571; <https://doi.org/10.3390/ani11020571>
- [20] Abdoli R, Ziaeddin Mirhoseini S, Ghavi Hossein-Zadeh N, Zamani P. Screening for causative mutations of major prolificacy genes in Iranian fat-tailed sheep. *Int J Fertil Steril* 2018; 12(1):51–5; <https://doi.org/10.22074/ijfs.2018.5247>
- [21] Hanrahan JP, Gregan SM, Mulsant P, Mullen M, Davis GH, Powell R, et al. Mutations in the genes for oocyte-derived growth factors GDF9 and BMP15 are associated with both increased ovulation rate and sterility in Cambridge and Belclare sheep (*Ovis aries*). *Biol Reprod* 2004; 70(4):900–9; <https://doi.org/10.1095/biolreprod.103.023093>
- [22] Abdelgadir AZ, Musa LMA, Jawasreh KI, Saleem AO, El-Hag F, Ahmed MA. G1 point mutation in growth differentiation factor 9 gene affects litter size in Sudanese desert sheep. *Vet World* 2021; 14(1):104–12; <https://doi.org/10.14202/vetworld.2021.104-112>
- [23] Aboelhassan DM, Darwish AM, Ali NI, Ghaly IS, Farag IM. A study on mutation points of GDF9 gene and their association with prolificacy in Egyptian small ruminants. *J Genet Eng Biotechnol* 2021; 19(1):85; <https://doi.org/10.1186/s43141-021-00181-8>
- [24] Gorlov IF, Kolosov YA, Shirokova NV, Getmantseva LV, Slozhenkina MI, Mosolova NI, et al. GDF9 gene polymorphism and its association with litter size in two Russian sheep breeds. *Rend Fis Acc Lincei* 2018; 29:61–6; <https://doi.org/10.1007/s12210-017-0659-2>
- [25] Moradband F, Rahimi G, Gholizadeh M. Association of polymorphisms in fecundity genes of GDF9, BMP15 and BMP15-1B with litter size in Iranian Baluchi sheep. *Asian Australas J Anim Sci* 2011; 24(9):1179–83; <https://doi.org/10.5713/ajas.2011.10453>
- [26] Rezaei V, Esmailizadeh A, Mehrgardi AA, Qanatqestani MD. Allelic polymorphism in exon 1 of GDF9 and exon 2 of BMP15 genes and its impact on litter size at lambing in Iran-Black sheep. *J Livest Sci Technol* 2020; 8(2):57–65; <https://doi.org/10.22103/jlst.2020.15007.1299>
- [27] Roy J, Polley S, De S, Mukherjee A, Batabyal S, Pan S, et al. Polymorphism of fecundity genes (FecB, FecX, and FecG) in the Indian Bonpala sheep. *Anim Biotechnol* 2011; 22(3):151–62; <https://doi.org/10.1080/10495398.2011.589239>
- [28] Jawasreh KIZ, Jadallah R, Al-Massad MM. Association between the prolificacy of Romanov sheep breed and fecundity gene, growth differentiation factor 9 gene and prolactin gene genotypes. *Malays Appl Biol* 2017; 46(2):1–8.
- [29] Javanmard A, Azadzadeh N, Esmailizadeh AK. Mutations in bone morphogenetic protein 15 and growth differentiation factor 9 genes are associated with increased litter size in fat-tailed sheep breeds. *Vet Res Commun* 2011; 35(3):157–67; <https://doi.org/10.1007/s11259-011-9467-9>
- [30] Abdoli R, Zamani P, Deljou A, Rezvan H. Association of BMPR-1B and GDF9 genes polymorphisms and secondary protein structure changes with reproduction traits in Mehraban ewes. *Gene* 2013; 524(2):296–303; <https://doi.org/10.1016/j.gene.2013.03.133>
- [31] Eghbalsaied S, Khorasgani FR, Amini HR, Farahi M, Davari M, Pirali A, et al. Variant GDF9 mRNA is likely not the main cause



- of larger litter size in Iranian Lori-Bakhtyari, Shal, Ghezel, and Afshari sheep breeds. *Arch Anim Breed* 2017; 60(2):119–29; <https://doi.org/10.5194/aab-60-119-2017>
- [32] Liandris E, Kominakis A, Andreadou M, Kapeoldassi K, Chadio S, Tsiligianni TH, et al. Associations between single nucleotide polymorphisms of GDF9 and BMP15 genes and litter size in two dairy sheep breeds of Greece. *Small Rumin Res* 2012; 107(1):16–21; <https://doi.org/10.1016/j.smallrumres.2012.04.004>
- [33] Pineda R, Montes D, Hernandez D. Association of the polymorphisms FecXR, FecGH, and FecGI and non-genetic factors that affect the prolificacy of Colombian Creole sheep. *Indian J Sci Tech* 2018; 11(17):1–7; <https://doi.org/10.17485/ijst/2018/v11i17/122374>
- [34] Talebi R, Ahmadi A, Afraz F, Sarry J, Woloszyn F, Fabre S. Detection of single nucleotide polymorphisms at major prolificacy genes in the Mehraban sheep and association with litter size. *Ann Anim Sci* 2018; 18(3):685–98; <https://doi.org/10.2478/aoas-2018-0014>
- [35] Lee YH. An overview of meta-analysis for clinicians. *Korean J Intern Med* 2018; 33(2):277–83; <https://doi.org/10.3904/kjim.2016.195>
- [36] Hoffman JIE. Meta-analysis. In: *Biostatistics for medical and biomedical practitioners*. Elsevier, Amsterdam, The Netherlands, pp 621–9, 2019.
- [37] Haidich AB. Meta-analysis in medical research. *Hippokratia* 2010; 14(Suppl 1):29–37.
- [38] Chong Y, Liu G, Jiang X. Effect of BMP15 gene on litter size of sheep in China: a meta-analysis. *Anim Reprod Sci* 2019; 210:106175; <https://doi.org/10.1016/j.anireprosci.2019.106175>
- [39] Zergani E, Rashidi A, Rostamzadeh J, Razmkabir M, Tetens J. Meta-analysis of association between c.963A>G single-nucleotide polymorphism on BMP15 gene and litter size in goats. *Arch Anim Breed* 2022; 65(3):309–18; <https://doi.org/10.5194/aab-65-309-2022>
- [40] Mahmoudi P, Rashidi A, Rostamzadeh J, Razmkabir M. Association between c.1189G>A single nucleotide polymorphism of GDF9 gene and litter size in goats: a meta-analysis. *Anim Reprod Sci* 2019; 209:106140; <https://doi.org/10.1016/j.anireprosci.2019.106140>
- [41] Page MJ, McKenzie JE, Bossuyt PM, Boutron I, Hoffmann TC, Mulrow CD, et al. The PRISMA 2020 statement: an updated guideline for reporting systematic reviews. *BMJ* 2021; 372:n71; <https://doi.org/10.1136/bmj.n71>
- [42] Irawan A, Hidayat C, Jayanegara A, Ratriyanto A. Essential oils as growth-promoting additives on performance, nutrient digestibility, cecal microbes, and serum metabolites of broiler chickens: a meta-analysis. *Anim Biosci* 2021; 34(9):1499–513; <https://doi.org/10.5713/ab.20.0668>
- [43] Higgins JPT, Green S. *Cochrane handbook for systematic reviews of interventions version 5.1.0*. London, UK, 2011. Available via <https://www.handbook.cochrane.org>; Stand: 15.01.2023
- [44] Balduzzi S, Rucker G, Schwarzer G. How to perform a meta-analysis with R: a practical tutorial. *Evidence-Based Mental Health* 2019; 22(4):153–60.
- [45] Higgins JP, Thompson SG, Deeks JJ, Altman DG. Measuring inconsistency in meta-analyses. *BMJ* 2003; 327(7414):557–60; <https://doi.org/10.1136/bmj.327.7414.557>
- [46] Lin L, Chu H. Quantifying publication bias in meta-analysis. *Biometrics* 2018; 74(3):785–94; <https://doi.org/10.1111/biom.12817>
- [47] Al-Mutar H, Younis L. Investigation effect of the point mutation in oocyte particular growth differentiation factor 9 gene on sterility and fertility in local Awassi sheep. *Arch Razi Inst* 2020; 75(1):101–8; <https://doi.org/10.22092/ari.2018.122232.1220>
- [48] Gurevitch J, Koricheva J, Nakagawa S, Stewart G. Meta-analysis and the science of research synthesis. *Nature* 2018; 555(7695):175–82; <https://doi.org/10.1038/nature25753>
- [49] Bangar YC, Magotra A, Chauhan A, Yadav AS. Genetic polymorphisms of kappa casein gene and its association with milk and composition traits in cows: an updated meta-analysis. *Meta Gene* 2021; 30:100948; <https://doi.org/10.1016/j.mgene.2021.100948>
- [50] Hossain F, Suma SA, Bhuiyan MSA. Association of GDF9 gene polymorphism with litter size in Indigenous sheep of Bangladesh. *Res Agric Livest Fish* 2020; 7(2):283–292; <https://doi.org/10.3329/ralf.v7i2.48871>